PONNAIYAH RAMAJAYAM INSTITUTE OF SCIENCE & TECHNOLOGY (PRIST)

Declared as DEEMED-TO-BE-UNIVERSITY
U/s 3 of UGC Act, 1956

SCHOOL OF ARTS AND SCIENCE

DEPARTMENT OF BIOCHEMISTRY

M.Sc., BIOCHEMISTRY CURRICULUM

23 REGULATION

PONNAIYAH RAMAJAYAM INSTITUTE OF SCIENCE & TECHNOLOGY (PRIST)

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MSc Biochemistry
2023 regulation

Programme Outcomes:

PO1. To make students understand the importance of biochemistry as a subject that deals with life processes, as well as the concepts, theories and experimental approaches followed in biochemistry, in order to pursue a research career, either in an industry or academic setting.

PO2. To develop analytical and problem-solving skills

PO3. To create an awareness among the students on the interconnection between the interdisciplinary areas of biochemistry.

PO4. To give the necessary practical skills required for biochemical techniques and analysis.

PO5. To develop ~~a~~ communication and writing skills in students.

PO6. To develop leadership and teamwork skills

PO7. To emphasiz**e** the importance of good academic and work ethics and their social implications.

PO8. To emphasize the importance of continuous learning and to promote lifelong learning and career development.

PO9. To teach students how to retrieve information from a variety of sources, including libraries, databases and the internet.

PO10. To teach students to identify, design and execute a research problem, analyze and interpret data and learn time and resource management.

PROGRAMME EDUCATIONAL OBJECTIVE (PEOs)

- **PEO 1**. The course aims to impart advanced and in depth understanding on all the human physiological and pathological state. To understand the molecular process and their perturbation during disease.

- **PEO 2**. The programme covers various aspects of Biomolecule estimation and regulation to ascertain health and disease state. metabolic pathways alterations along with their regulation at the replication, transcriptional, translational, and post-translational levels including by studying DNA, RNA and protein molecules, immunology, endocrinology, advancements in rDNA technologies to circumvent genetic disorders.

- **PEO 3**. Further to enrich research understanding various genomic, proteomic and bioinformatics tools are added. Animal cell culture, IPR, Biostatistics, research methodology, clinical research and Plant tissue culture are offered as elective papers to get specialized in a specific area. The final semester is devoted exclusively to enrich the students to address specific research objective.

- **PEO 4.:** Understand the role of citizen to maintain sustainable environment and encourage Eco-friendly initiatives.

- **PEO 5.:** Acquire the ability to engage in independent and life-long learning in the broadest context of health and disease.

Programme Specific Outcomes:

On successful completion of this course, students should be able to:

PSO1. Understand the principles and methods of various techniques in Biochemistry, Immunology, Microbiology, Enzyme kinetics and Molecular Cell Biology. Based on their understanding, the students may would be able to design and execute experiments during their final semester project, and further research programs.

PSO2. Insight on the structure-function relationship of biomolecules, their synthesis and breakdown, the regulation of these pathways, and their importance in terms of clinical correlation. Students will also acquire knowledge of the principles of nutritional biochemistry and also understand diseases and their prevention.

PSO3. To understand the concepts of cellular signal transduction pathways and the association of aberrant signal processes with various diseases. Acquire insight into the immune system and its responses, and use this knowledge in the processes of immunization, vaccine development, transplantation and organ rejection.

PSO4. To visualize and appreciate the central dogma of molecular biology, regulation of gene expression, molecular techniques used in rDNA technology, gene knock-out and knock-in techniques.

PSO5. To create awareness in students about the importance of good laboratory practices and the importance of ethical and social responsibilities of a researcher. Teach them how to review literature and the art of designing and executing experiments independently and also work as a part of a team.

C1- Basics of Biochemistry
C2- Biochemical and Molecular Biology Techniques
C3- Physiology and Cell Biology
C4- Biomolecules and Biochemical Techniques Lab
C5- Enzymology
C6- Cellular Metabolism
C7- Clinical Biochemistry
C8- Enzymology , Microbiology and Cell Biology Lab
C9- Nutritional Biochemistry
C10- Industrial Microbiology
C11- Participation in Bounded Research
C12- Industrial Microbiology
C13-Industrial Microbiology
C14- Molecular Biology
C15- Clinical Biochemistry Lab
C16- Gene Editing, Cell and Gene therapy
C17- Molecular basis of disease and therapeutic strategies
C18- Pharmaceutical Biochemistry
C19- Biochemical Toxicology
C20- Project Work
C21-Discipline Specific Elective I-V

M. Sc., Curriculum Mapping
Programme Educational Objectives vs Programme Outcome

POs	1	2	3	4	5	6	7	8	9	10
PEO I	*		*			*	*	*		
PEO II	*		*	*	*	*	*	*		*
PEO III	*	*	*	*	*		*		*	
PEO IV		*		*		*		*		*
PEO V	*		*		*		*		*	

M. Sc., Curriculum Mapping

Programme Outcome vs Course Outcome

Programme Outcome-PO Course Outcome-CO	PO1	PO2	PO3	PO4	PO5
CO1	*	*	*	*	*
CO2	*	*		*	*
CO3	*	*	*	*	*
CO4	*	*	*		
CO5	*	*	*	*	*
CO6	*	*	*	*	*
CO7				*	*
CO8	*	*	*	*	*
CO9				*	*
CO10	*	*	*	*	*
CO11			*	*	*

CO12	*	*	*	*	*
CO13	*	*	*	*	*
CO14				*	*
CO15	*	*	*	*	*
CO16				*	*
CO17	*	*	*	*	*
CO18		*	*	*	*
CO19	*	*	*	*	*
CO20	*	*	*		
CO21	*	*	*		

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Course Structure

MSc Biochemistry

2023 regulation

Course Code	Course Title	L	T	P	C
	SEMESTER I				
23215AEC11	Basics of Biochemistry	5	1	0	4
23215AEC12	Biochemical and Molecular Biology Techniques	5	1	0	4
23215AEC13	Physiology and Cell Biology	5	1	0	4
23215SEC14L	Biomolecules and Biochemical Techniques Lab	0	0	4	4
23215DSC15_ _	Discipline Specific Elective I	5	1	0	3
23215RMC16	Research Methodology	2	-	-	2
	Total	22	4	4	21
	SEMESTER II				
23215AEC21	Enzymology	4	1	0	4
23215AEC22	Cellular Metabolism	4	1	0	4
23215AEC23	Clinical Biochemistry	4	1	0	4
23215SEC24L	Enzymology , Microbiology and Cell Biology Lab	0	0	4	4
23215DSC25_	Discipline Specific Elective II	4	1	0	3
23215SEC26	Nutritional Biochemistry	4	0	0	3
23215BRC27	Participation in bounded Research	2	0	0	2
23215SEC28	Internship-[Clinical Laboratory]	-	-	-	2
	Total	22	4	4	26
	SEMESTER III				
23215AEC31	Industrial Microbiology	5	1	0	4
23215AEC32	Molecular Biology	4	1	0	4
23215AEC33	Gene Editing, Cell and Gene therapy	4	1	0	4
23215SEC34L	Clinical Biochemistry Lab	0	0	4	4
23215DSC35_	Discipline Specific Elective III	4	1	0	3

23215SEC36	Molecular basis of disease and therapeutic strategies	4	1	0	3
23215SEC37	Industrial Visit – Biotech	-	-	-	2
	Total	21	5	4	24
	SEMESTER IV				
23215AEC41	Pharmaceutical Biochemistry	4	1	0	4
23215AEC42	Biochemical Toxicology	4	1	0	4
23215PRW43	Project and viva	0	0	10	4
23215DSC44_	Discipline Specific Elective IV	4	1	0	3
23215DSC45_	Discipline Specific Elective v	4	1	0	3
23215SEC46	Industrial Visit – Pharma or Food Processing	-	-	-	2
	Total	16	4	10	20
	Total Credits for the Programme				91

Discipline specific Electives

Semester	Discipline specific Elective Courses-I
I	a)20215DSC15A– Microbiology & Immunology b) 20215DSC15B– Endocrinology
	Discipline specific Elective Courses-II
II	a)2015DSC25A- Energy and drug metabolism b)20215 DSC25B- Neuro Biochemistry
	Discipline specific Elective Courses-III
III	a)20215DSC34A- Biostatistics and Data Science b)20215DSC34B- Immunology
	Discipline specific Elective Courses-IV
IV	a) 20215DSC44A – Biosafety, Lab Safety and IPR b) 20215DSC44B – Plant Biochemistry
	Discipline specific Elective Courses-V
IV	a) 20215DSC45A -Developmental Biology b) 20215DSC45B -Cancer Biology

Credit Distribution:

Sem	**AEC**	**SEC**	**DSC**	**RSB Courses**	**Others**	**Total**
I	12	4	3	2	-	21
II	12	7	3	2	2	26
III	12	7	3	-	2	24
IV	8	-	6	4	2	20
Total	**44**	**18**	**15**	**08**	**06**	**91**

Course Code	Course name	L	T	P	C
23215AEC11	**BASICS OF BIOCHEMISTRY**	5	1	0	4

Pre-requisites, ifany: Basic Knowledge of Biochemistry and Biomolecules

Course Objectives

The main objectives of this course are to:

1. Students will be introduced to the structure of biomolecules.
2. The significance of carbohydrates in biological processes will be understood.
3. The structure, properties and biological significance of lipids in the biological system will be studied
4. Students will learn about the concepts of protein structure and their significance in biological processes and creatively comprehend the role of membrane components with theirbiological significance.

Students will gain knowledge about the structures and functional roles of nucleic acids in the biological system

Course Outcomes

On successful completion of the course, the students should be able to:

CO1 : Explain the chemical structure and functions of carbohydrates.(K1, K2)

CO2: Using the knowledge of lipid structure and function, explain how it plays a role in

Signalling pathways (K3,K4)

CO3: Describe the various levels of structural organisation of proteins and the role of proteins in biological system (K4, K5)

CO4: Apply the knowledge of proteins in cell-cellinteractions.(K3,K4)

CO5. Applying the knowledge of nucleic acid sequencing in research and diagnosis (K2, K3,K4)

UNIT I

Carbohydrates- Classification, structure (configurations and conformations, anomeric forms), function and properties of monosaccharides, mutarotation, Disaccharides and oligosaccharides with suitable examples . Polysaccharides - Homopolysaccharides (starch, glycogen, cellulose, inulin, dextrin, agar, pectin, dextran). Heteropolysaccharides - Glycosaminoglycans– source, structure, functions of hyaluronic acid, chondroitin sulphates, heparin, keratan sulphate,. Glycoproteins - proteoglycans. O- Linked and N-linked glycoproteins. Biological significance of glycan. Blood group polysaccharides. Bacterial cell wall (peptidoglycans, teichoic acid) and plant cell wall carbohydrates.

UNIT II: Lipids – Classification of lipids, structure, properties and functions of fatty acids, triacylglycerols, phospholipids, glycolipids, sphingolipids and steroids – Biological importance. Eicosanoids- classification, structure and functions of prostaglandins, thromboxanes, leukotrienes. Lipoproteins – Classification ,structure, transport (endogenous and exogenous Pathway) and their biological significance.

UNIT III: Overview of Amino acids - classification, structure and properties of amino acids, Biological role.Non Protein aminoacids and their biological significance .Proteins –

classification based on composition, structure and functions. Primary, secondary, super secondary (motifs) (Helix-turn –helix, helix-loop-helix, Beta-alpha-beta motif, Rosemann Rossmann fold , Greek key),tertiary and quaternary structure of proteins. Structural characteristics of collagen and hemoglobin. Determination of amino acid sequence.Chemical synthesis of a peptide, Forces involved in stabilization of protein structure. Ramachandran plot. Folding of proteins. Molecular chaperons – Hsp 70 and Hsp 90 - biological role.

UNIT IV: Membrane Proteins - Types and their significance. Cytoskeleton proteins - actin , tubulin , intermediate filaments . Biological role of cytoskeletal proteins. Membrane structure-fluid mosaic model

UNIT V: Nucleic acids – types and forms (A, B, C and Z) of DNA. Watson-Crick model-Primary, secondary and tertiary structures of DNA. Triple helix and quadruplex DNA. Mitochondrial and chloroplast DNA. DNA supercoiling (calculation of Writhe, linking and twist number). Determination of nucleic acid sequences by Maxam Gilbert and Sanger's methods. Forces stabilizing nucleic acid structure. Properties of DNA and RNA. C-value, C-value paradox, Cot curve. Structure and role of nucleotides in cellular communications. Major and minor classes of RNA, their structure and biological functions.

Self-Study

1. Classification of Sugars
2. Nutritional classifcation of fatty acids

Recommended Texts

1. David L.Nelson and Michael M.Cox (2012) Lehninger Principles of Biochemistry (6th ed) W.H. Freeman.

2. Voet.D & Voet. J.G (2010) Biochemistry , (4th ed), JohnWiley & Sons, Inc.

3. Metzler D.E (2003). The chemical reactions of livingcells (2nd ed), Academic Press.

4. Zubay G.L (1999) Biochemistry , (4th ed), Mc Grew-Hill.

5. Lubert Stryer (2010) Biochemistry,(7th ed), W.H.Freeman

Satyanarayan,U (2014) Biochemistry (4th ed), ArunabhaSen Books & Allied (P) Ltd, Kolkata.

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse(K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create (K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	L	M	S	M	M	M	S	M	M
CO 2	S	M	L	S	M	M	M	S	M	M
CO 3	S	M	M	S	S	M	L	S	M	M
CO 4	S	M	M	S	M	M	M	S	M	M
CO 5	S	S	M	S	S	M	M	S	M	M

S- Strong M-Medium L-Low

Course Code	Course name	L	T	P	C
23215AEC12	**BIOCHEMICAL AND MOLECULAR BIOLOGY TECHNIQUES**	5	1	0	4

Pre-requisites, if any: Comprehensive Knowledge of Tools of Biochemistry/Molecular Biology

Course Objectives

Biochemical techniques combine various inter-disciplinary methods in biological research and the course aims to provide students with the following objectives:

1. To understand the various techniques used in biochemical investigation and microscopy.
2. To explain chromatographic techniques.\ and their applications
3. To explain electrophoretic techniques.
4. To comprehend the spectroscopic techniques and demonstrate their applications in biochemical investigations.
5. To acquire knowledge of radio labelling techniques and centrifugation.

Course Outcomes

After completion of the course, the students should be able to:

CO1. Attain good knowledge in modern used in biochemical investigation and microscopy and apply the experimental protocols to plan and carry out simple investigations in biological research. (K1, K5)

CO2. Demonstrate knowledge to implement the theoretical basis of chromatography in upcoming practical course work. (K3, K5)

CO3. Demonstrate knowledge to implement the theoretical basis of

electrophoretic techniques in research work. (K3, K5)

CO4. Tackle more advanced and specialized spectroscopic techniques that are pertinent to research. (K1, K2 & K5)

CO5. Tackle more advanced and specialized radioisotope and centrifugation techniques that are pertinent to research work. (K1, K2 & K5)

Units I

General approaches to biochemical investigation, cell culture techniques and microscopic techniques. Organ and tissue slice technique, cell distribution and homogenization techniques, cell sorting, and cell counting, tissue Culture techniques. Cryopreservation, Biosensors- principle and applications. Principle, working and applications of light microscope, dark field, phase contrast and fluorescent microscope. Electron microscope- Principle, instrumentation of TEM and SEM, Specimen preparation and applications-shadow casting, negative staining and freeze fracturing.

Unit **II**

Chromatographic Techniques:

Basic principles of chromatography- adsorption and partition techniques. Chiral Chromatography and counter current Chromatography. Adsorption Chromatography – Hydroxy apatite chromatography and hydrophobic interaction Chromatography. Affinity chromatography.Gas liquid chromatography- principle, instrumentation, column development, detectors and applications. Low pressure column chromatography – principle, instrumentation, column packing, detection, quantitation and column efficiency, High pressure liquid chromatography- principle, instrumentation, delivery pump, sample injection unit, column packing, development, detection and application. Reverse HPLC, capillary electro chromatography and perfusion chromatography.

Unit III **Electrophoretic Techniques:**

General principles of electrophoresis, supporting medium, factors affecting electrophoresis, Isoelectric focusing-principle, ampholyte, development of pH gradient and application. PAGE-gel casting-horizontal, vertical, slab gels, sample application, detection-staining using CBB, silver, fluorescent stains. SDS PAGE-principle and application in molecular weight determination principle of disc gel electrophoresis ,2D PAGE. Electrophoresis of nucleic acids-agarose gel electrophoresis of DNA, pulsed field gel electrophoresis- principle, apparatus, application. Electrophoresis of RNA, curve. Microchip electrophoresis and 2D electrophoresis, Capillary electrophoresis.

Unit IV **Spectroscopic techniques:**

Basic laws of light absorption- principle, instrumentation and applications of UV-Visible, IR, ESR, NMR, Mass spectroscopy, Turbidimetry and Nephelometry. Luminometry (Luciferase system, chemiluminescence). X - ray diffraction. Atomic absorption spectroscopy - principle and applications - Determination of trace elements

Unit V **Radiolabeling Techniques and Centrifugation:**

Nature of radioactivity-detection and measurement of radioactivity, methods based upon ionisation (GM counter) and excitation (scintillation counter), autoradiography and applications of radioactive isotopes, Biological hazards of radiation and safety measures in handling radioactive isotopes.

Basic principles of Centrifugation. Preparative ultracentrifugation - Differential centrifugation, Density gradient centrifugation. Analytical ultracentrifugation - Molecular weight determination.

Reading List

(Print and Online)

Principles and techniques of biochemistry and molecular biology:

https://www.kau.edu.sa/Files/0017514/Subjects/principals%20and%20techniques%20of%20biochemistry%20and%20molecular%20biology%207th%20ed%

Self-Study	1.	Types of rotors
	2.	Colorimetry – principle and applications

Recommended Texts

1.Keith Wilson , John Walker (2010) Principles and Techniques of Biochemistry and Molecular Biology (7th ed) Cambridge University Press

2.David Sheehan (2009), Physical Biochemistry: Principles and Applications (2nd ed), Wiley-Blackwell

3.David M. Freifelder (1982) Physical Biochemistry: Applications to Biochemistry and Molecular Biology,W.H.Freeman

4.Rodney F.Boyer (2012), Biochemistry Laboratory: Modern Theory and techniques,(2nd ed),Prentice Hall

5.Kaloch Rajan (2011), Analytical techniques in Biochemistry and Molecular Biology, Springer

6. Segel I.H (1976) Biochemical Calculations (2nd ed),John Wiley and Sons

7. Robyt JF (2015) Biochemical techniques: Theory and Practice (1st ed), CBS Publishers & Distributors

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse(K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create (K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	**S**	**L**	**M**	**S**	**S**	**L**	**L**	**S**	**S**	**M**
CO 2	**S**	**M**	**M**	**S**	**M**	**L**	**M**	**S**	**S**	**L**
CO 3	**S**	**M**	**L**	**S**	**M**	**M**	**M**	**S**	**M**	**L**
CO 4	**S**	**S**	**L**	**S**	**S**	**M**	**M**	**S**	**M**	**M**
CO 5	**S**	**S**	**M**	**S**	**M**	**M**	**M**	**S**	**M**	**M**

S-Strong MS-Strong M-Medium L-Low

Course Code	Course name	L	T	P	C
23215AEC13	**PHYSIOLOGY AND CELL BIOLOGY**	5	1	0	4

Pre-requisites, if any:	Anatomy, Cells and Biological Compounds
Course Objectives	To understand the functions and activities of organs, tissues or cells and of physical and chemical phenomena involved in the human body
Course Outcomes	After completion of the course, the students should be able to: **CO1.** specifically understand the biological and chemical processes within a human cell (K1, K2, K5, K6) **CO2.** identify and prevent diseases (K2, K3, K4, k5, K6) **CO3.** understand defects in digestion, nutritional deficiencies and intolerances, and gastrointestinal pathologies (K1, K2, K3, K4, K5, K6) **CO4.** identify general characteristics in individuals with imbalances of acid- base, fluid and electrolytes.(K1 , K2 , K3 , K4, K5, K6) **CO5.** process the mechanism: the transmission of biochemical information between cell membrane and nucleus. (K1, K2, K5)
Unit I	Major classes of cell junctions- anchoring, tight and gap junctions. Major families of cell adhesion molecules (CAMs)- cadherins, integrins. Types

of tissues. Epithelium- organisation and types. The basement membrane. Cell cycle- mitosis and meiosis, Cell cycle-phases and regulation. Cell death mechanisms- an overview-apoptosis, necrosis.

Unit II Reproductive system- sexual differentiation and development; sperm transport, sperm capacitation, semen analyses and Acrosome reaction. Clinical relevance of female reproductive physiology- menstrual cycle, pregnancy and menopause. Fertilisation and infertility issues.

Unit III Digestive system- structure and functions of different components of digestive system, digestion and absorption of carbohydrates, lipids and proteins, role of bile salts in digestion and absorption, mechanism of HCl formation in stomach, role of various enzymes and hormones involved in digestive system. Composition of blood, lymph and CSF. Blood cells - WBC, RBC and energy metabolism of RBC, Blood clotting mechanism and blood groups- ABO and Rhesus system.

Unit IV Respiratory system-Gaseous transport and acid-base homeostasis. Mechanism of the movement of O2 and CO2 through lungs, arterial and venous circulation. Bohr effect, oxygen and carbon dioxide binding haemoglobin. pH maintenance by cellular and intracellular proteins. Phosphate and bicarbonate buffers, Metabolic acidosis and alkalosis. Respiratory acidosis and alkalosis. Regulation of fluid and electrolyte balance.

Unit V Sensory transduction, Nerve impulse transmission- nerve cells, synapses, reflex arc structure, resting membrane potential, Nernst equation, action potential, voltage gated ion-channels, impulse transmission, neurotransmission, neurotransmitter receptors, synaptosomes, synaptotagmin, rod and cone cells in the retina, changes in the visual cycle, photochemical reaction and regulation of rhodopsin, odour receptors, learning and memory. Chemistry of muscle contraction – actin and myosin filaments, theories involved in muscle contraction, mechanism

	of muscle contraction, energy sources for muscle contraction.
VI	Hormones – Classification, Biosynthesis, circulation in blood, modification and degradation. Mechanism of hormone action, Target cell concept. Hormones of Hypothalamus, pituitary, Pancreatic, thyroid & parathyroid, adrenal and gonadal hormones. Synthesis, secretion, physiological actions and feedback regulation of synthesis.
Reading List (Print and online)	https://www.genome.gov/genetics-glossary/Cell-Cycle https://my.clevelandclinic.org/health/diseases/16083-infertility-causes https://www.webmd.com/heartburn-gerd/reflux-disease https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5760509/ https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3249628/
Self-Study	1. Variation in cell differentiation and progression 2. Lesch Nyhan syndrome , orotic aciduria and GERD
Recommended Texts	1. Karp, G. (2010). Cell and Molecular Biology: Concepts and Experiments (6th ed). John Wiley & Sons. Inc. 2. Bruce Alberts and Dennis Bray (2013),Essential Cell Biology,(4th ed),Garland Science. 3. De Robertis, E.D.P. and De Robertis, E.M.F. (2010). Cell and Molecular Biology.(8th ed). Lippincott Williams and Wilkins, Philadelphia. 4. Cooper, G.M. and Hausman, R.E. (2009). The Cell: A Molecular Approach. (5th ed). Sunderland, Mass. Sinauer Associates, Inc. 5. Wayne M. Baker (2008) the World of the Cell. (7th ed). Pearson Benjamin Cummings Publishing, San Francisco. Cell Biology 6. John E. Hall (2010). Guyton and Hall Textbook of Medical Physiology (12th ed), Saunders 7. Harrison's Endocrinology by J. Larry Jameson Series: Harrison's Specialty, 19th Edition Publisher: McGraw-Hill, Year: 2016.

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) -Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations. Discussion

Mapping with Programme Outcomes:

	PO 1	**PO 2**	**PO 3**	**PO 4**	**PO 5**	**PO 6**	**PO 7**	**PO 8**	**PO 9**	**PO 10**
CO 1	**S**	**S**	**S**	**S**	**S**	**M**	**S**	**S**	**S**	**M**
CO 2	**S**	**S**	**S**	**S**	**S**	**L**	**S**	**S**	**S**	**M**
CO 3	**S**	**S**	**S**	**S**	**S**	**M**	**M**	**S**	**S**	**M**
CO 4	**S**	**S**	**S**	**S**	**S**	**M**	**M**	**S**	**S**	**M**
CO 5	**M**	**S**	**L**	**S**	**S**	**L**	**M**	**M**	**L**	**L**

S-Strong M-Medium L-Low

Course Code	Course name	L	T	P	C
23215SEC14L	Biomolecules and Biochemical Techniques Lab	**0**	**0**	**4**	**4**

Pre-requisites Knowledge on basic principles, Instrumentation of Biochemical techniques and metabolic reactions

Course Objectives

1. To instill skill in students enabling them to apprehend the wider knowledge about principles and techniques to be employed for the biomolecules under investigation.

2.To inculcate the knowledge of various isolation and purification techniques of macromolecules like DNA, RNA, Glycogen and Starch,

3. To perform colorimetric estimations to quantify important metabolites like lactate and tryptophan and minerals like calcium and iron from various sources.

4. To achieve training in subcellular fractionation and to identify them by markers.

5 To achieve training in various chromatographic techniques.

6. To perform the isolation and identification of the organelles of a cell using differential centrifugation.

7. To perform phytochemical screening and quantification enabling them to give an insight on phytochemicals this will be useful for future research.

Course Outcomes **On successful completion of this course, students should be able to:**

After completion of the course, the students should be able to:

CO1. The student will be able to acquire knowledge and skill in the techniques used in the isolation, purification and estimation of different biomolecules that are widely employed in research (K1, K2, K4)

CO2. The students will get acquainted with Principle, Instrumentation and method of Performing UV absorption studies of DNA, Protein and interpreting the alteration occurred during the process of denaturation (K1,K2, K 3, K4).

CO3. The student will be fine-tune in handling the instruments like colorimeter, spectrophotometer and will be able to estimate the biomolecules and minerals from the given samples (K1,K2,K4,)

CO4. The student, in addition to acquiring skill in performing various biochemical techniques can also learn to detect presence of phytochemicals and quantify them in the plant sample. (K1,K2,K3,K4 & K6)

CO5. The students will develop skill in analytical techniques like subcellular fractionation, Paper, Column and Thin layer Chromatography and the group experiments will enable them to build learning skills like team work, Problem solving, Communication ability. (K1, K2,K3,K4 & K6)

Units I Biochemical studies and estimation of macromolecules

1. Isolation and estimation of glycogen from liver.
2. Isolation and estimation of DNA from animal tissue.
3. Isolation and estimation of RNA from yeast.
4. Purification of Polysaccharides –Starch and assessment of its purity

Units II UV absorption

1. Denaturation of DNA and absorption studies at 260nm.
2. Denaturation of Protein and absorption studies at 280nm.

Units III Colorimetric estimations

1. Estimation of Pyruvate
2. Estimation of tryptophan.

Units IV Estimation of minerals

1. Estimation of calcium
2. Estimation of iron

Units V	Plant Biochemistry 1. Qualitative analysis Phytochemical screening 2.Estimation of Flavonoids -Quantitative analysis
Units VI	Group Experiments 1.Fractionation of sub-cellular organelles by differential centrifugation-Mitochondria and nucleus 2. Identification of the separated sub-cellular fractions using marker enzymes (any one) 3.Separation of identification of lipids by thin layer chromatography.. 4.Separation of plant pigments from leaves by column chromatography 5. Identification of Sugars by Paper Chromatography 6.Identification of Amino acids by Paper Chromatography
Reading List (Print and Online)	1.https://www.researchgate.net/publication/313745155_Practical_Biochemistry_A_Student_Companion 2.https://doi.org/10.1186/s13020-018-0177-x 3.https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5368116/ 4.https://www.life.illinois.edu/biochem/455/Lab%20exercises/2Photometry/spectrophotometry.pdf 5.https://ijpsr.com/bft-article/determination-of-total-flavonoid-and-phenol-content-in-mimusops-elengi-linn/?view=fulltext 6.https://skyfox.co/wp-content/uploads/2020/12/Practical-Manual-of-Biochemistry.pdf
Self-Study	1. Laboratory Safety Rules, Requirements and Regulations. 2. Preparation of standard solutions and reagent
Books Recommended	1. David Plummer (2001) An Introduction to Practical Biochemistry (3rd ed) McGraw Hill Education (India) Private Ltd 2. Jayaraman, J (2011),laboratory Manual in Biochemistry, New age publishers 3. Varley H (2006) Practical Clinical Biochemistry (6th ed) , CBS Publishers

4. O. Debiyi and F. A. Sofowora, (1978)"Phytochemical screening of medical plants," Iloyidia, vol. 3, pp. 234–246,
5. Prof. Sarin A. Chavhan, Prof. Sushilkumar A. Shinde (2019) A Guide to Chromatography Techniques Edition:1
6. Analytical techniques in Biochemistry and Molecular Biology; Katoch, Rajan. Springer (2011)

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	**PO 2**	**PO 3**	**PO 4**	**PO 5**	**PO 6**	**PO 7**	**PO 8**	**PO 9**	**PO 10**
CO 1	S	S	S	S	M	S	L	S	M	S
CO 2	S	S	S	S	M	S	L	S	M	S
CO 3	S	S	S	S	M	S	M	S	M	S
CO 4	S	S	S	S	S	S	S	S	S	S
CO 5	S	S	S	S	S	S	S	S	S	S

S-Strong M-Medium L-Low

Course Code	Course name	L	T	P	C
23215DSC15A	**MICROBIOLOGY & IMMUNOLOGY**	5	1	0	3

Pre-requisites, if any: The student should possess basic knowledge about microorganisms, types and their general characteristics. The students are also expected to possess basic understanding about the process of infection, immunological defence and pathological outcomes, if any.

Course Objectives

1. To appreciate the classification of microorganisms based on their structure, size and shape with an insight into the ancient scriptures about microbes.
2. To understand the role of microorganisms in environment and also to learn the culture conditions.
3. To recognize the possible contamination of foods by microorganisms, to learn about counteracting preservative measures and to know about probiotic nature of microorganisms.
4. To gain knowledge on pathogenic mediation by microorganisms and preventive measures as well.
5. To comprehend the features of antimicrobial agents, their mechanism of action along with the side effects and also to explore natural remedial measures against microbes.
6. To be able to exploit the various features of microorganisms for the beneficial industrial production.

Course Outcomes

After completion of the course, the students should be able to:

CO1. To classify (by both ancient and modern modes) different types of microorganisms and explain life cycle of the microbes (K1, K2 & K5)

CO2. To recognize the microorganisms involved in decay of foods and will be able to apply various counteracting measures. The students also will be able to relate the role of certain beneficial microbes in day-to- day's food consumption. (K1, K2 & K4)

CO3. To understand the common pathogenic bacterial and fungi that cause toxic effects and also will be able to employ curative measures. (K1 & K2)

CO4. To analyse various features of wide variety of antimicrobial agents along with their mode of action, in addition, being able to apprehend the valuable potentials of traditional and easily available herbs. (K2, K5 & K6)

CO5. To apply knowledge gained in production of industrially important products as both pharmaceutical and nutraceutical. (K2, K4 & K5)

Units I

Taxonomical classification - bacteria, viruses (DNA, RNA), algae, fungi and protozoa. Distribution and role of microorganisms in soil, water and air. Charaka's classification of microbes, lytic cycle and lysogeny. Types of culture media, isolation of pure culture, growth curve and the measurement of microbial growth.

Units II

Contamination and spoilage of foods – cereals, cereal products, fruits, vegetables, meat, fish, poultry, eggs, milk and milk products. General principles of traditional and modern methods of food preservation - Removal or inactivation of microorganisms, boiling, steaming, curing, pasteurization, cold processing, freeze drying, irradiation, vacuum packing, control of oxygen and enzymes. Microbes involved in preparation of fermented foods - cheese, yoghurt, curd, pickles, rice pan cake, appam, ragi porridge (கேழ்வரகு கூழ்) and bread.

Units III

Food poisoning- bacterial food poisoning, *Salmonella*, *Clostridium blotulinum* (botulism), *Staphylococcus aureus*, fungal food poisoning – aflatoxin, food infection – *Clostridium*, *Staphylococcus* and *Salmonella*. Pathogenic microorganisms, *E. coli, Pseudomonas, Klebsilla, Streptococcus, Haemophilus, & Mycobacterium*, causes, control, prevention, cure and safety. Food microbiological screening- Real time PCR, ELISA, Aerobic and anaerobic Plate Count, dye reduction method, anaerobic lactic acid bacteria, anaerobic sporeformers, Hazard analysis critical control point (HACCP)

Units IV	Antimicrobial chemotherapy, General characteristics of antimicrobial agents. Mechanism of action – sulfonamides, sulphones and PAS. Penicillin, streptomycin- spectra of activity, mode of administration, mode of action, adverse effects and sensitivity test., Antiviral and antiretroviral agents, Antiviral RNA interference, natural intervention (Natural immunomodulators routinely used in Indian medical philosophy).
Units V	Immune system- definition and properties. Cells of the immune system – neutrophils, eosinophils, basophils, mast cells, monocytes, macrophages, dendritic cells, natural killer cells, and lymphocytes (B cells and T cells). Lymphoid organs- Primary and Secondary; structure and functions. Antigens and Complement System: definition, properties- antigenicity and immunogenicity, antigenic determinants and haptens. Antigen - antibody interactions - molecular mechanism of binding. Affinity, avidity, valency, cross reactivity and multivalent binding. Immunoglobulins & Immune Response: Structure, classes and distribution of antibodies. Antibody diversity. Immune system in health & disease, Transplantation immunology- graft rejection and HLA antigens. Immunological techniques, Flow cytometry and its application.
Reading List (Print and Online)	https://www.ijam.co.in/index.php/ijam/article/view/1326 (Krumi (Microorganisms) in Ayurveda- a critical review) Virtual Lectures in Microbiology and Immunology, University of Rochester https://www.frontiersin.org/articles/10.3389/fphar.2020.578970/full#h9 https://www.frontiersin.org/articles/10.3389/fmicb.2018.02151/full https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7559905/
Self-Study	1. Microbial infections and gut microbiome with relevance to *tridoshas* 2. Microbial population and pH variations in different dairy products.
Recommended	1.Michael J.Pelczar Jr.(2001) Microbiology (5th ed), McGraw Hill Education (India) Private Limited

Texts

2. Frazier WC , Westhoff DC, Vanitha NM (2010) Food Microbiology (5th ed), McGraw Hill Education (India) Private Limited
3. Willey J and Sherwood L (2011) ,Prescott's Microbiology (8th ed) McGraw Hill Education (India)
4. Ananthanarayanan , Paniker and Arti Kapil (2013) Textbook of Microbiology (9th ed) Orient BlackSwan
5. Judy Owen , Jenni Punt Kuby (2013) ,Immunology (Kindt, Kuby Immunology) (7th ed) W. H. Freeman & Co
6. Brooks GF and Carroll KC (2013) Jawetz Melnick&Adelbergs Medical Microbiology,(26th ed) McGraw Hill Education
7. Greenwood D (2012) ,Medical Microbiology, Elsevier Health

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Observe, Explain.

Analyse (K4)- Finish procedure in stepwise manner, Differentiation between various ideas, Map knowledge

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion, Debating, Presentation

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	**S**	**L**	**S**	**S**	**S**	**S**	**M**	**S**	**S**	**S**
CO 2	**S**	**S**	**S**	**S**	**S**	**M**	**L**	**M**	**S**	**S**
CO 3	**S**	**M**	**M**	**S**	**M**	**M**	**M**	**M**	**L**	**M**
CO 4	**S**	**M**	**M**	**M**	**M**	**M**	**M**	**S**	**S**	**S**
CO 5	**S**	**L**	**S**	**S**	**M**	**L**	**L**	**S**	**S**	**S**

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC15B	Endocrinology	5	1	0	3

Aim:

To have a basic understanding of the endocrine system

Course Objective:

The specific objectives of the program are to train the fellows to:

- History and physical examination with emphasis on examination of the thyroid, breasts, penis, testes and female reproductive organs.
- Selection and interpretation of endocrine biochemical tests.

Course Outcomes:

- CO1Apply the knowledge from this course while working in medical laboratory to diagnose different hormone disorders
- CO2 Explain recent laboratory methods in diagnosis hormone disorders
- CO3 Knowledge and Understanding the synthesis of different endocrine gland hormones
- CO4 Ability to analyze and solve problems related to hormone tests
- CO5 To know the pathophysiology significance of the system with special reference to humans
- CO6-Understand the synthesis of various hormones by respective gland

UNIT: I

Hormones in general - definition – types of secretions – nature – classification synthesis and their role – Feed back control with specific examples Hormones action Proteins and Steroids – Cell Signaling in hormone action.

UNIT: II

Hypothalomo hypophysial axis - Hormones of hypothalamus and their role Structure of pituitary - Secretions - Physiology role -Pathophysiology Current status of pituitary as a master gland.

UNIT: III

Thyroid - Parathyroid - structure - hormones – synthesis – storage – releases- carrier proteins (eg. TBA and TBG) – Physiology role – Pathophysiology

UNIT: IV

Adrenal and Gonadal Hormones – Steroid biosynthesis – maintenance of cyclicity. Physiological role – Pathophysiology – Steroids in metabolism

UNIT: V

Gastro intestinal hormones – pancreas as an endocrine organ – secretions- functions – physiological role and pathophysiology other endocrine organs in vertebrate Insect and crustacean hormones – their role in growth and metamorphosis.

REFERENCE:

1. Text Book of endocrinology – Williams
2. Physiological review of Biochemistry – Harper and others
3. Endocrinology – Turner
4. Invertebrate reproduction – K K Nayar

Course Code	Course Title	L	T	P	C
20215RMC26	**Research Methodology**	**3**	**0**	**0**	**2**

AIM:

To create a basic appreciation towards research process and awareness of various research publication

COURSE OBJECTIVES:

- To understand the steps in research process and the suitable methods.
- To identify various research communications and their salient features
- To carry out basic literature survey using the common data-bases
- To give exposure to MATLAB platform for effective computational and graphic works required for quality research

COURSE OUTCOME:

- CO1 Understanding research questions and tools
- CO2 Experience in scientific writings
- CO3 Practice in various aspects of scientific publications
- CO4 Inculcation of research ethics

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UNIT I:

Selection of problem-stages in the execution of research: choosing a topic to publication-preparation of manuscript-report writing- format of journals – proof reading – sources of information: Journals, reviews, books, monographs, etc, Bibilograpy. Journal ; standard of research journals – Impact factor.

UNIT II:

Measures of dispersion: Universe and population – delimiting population – sampling method – random sampling, stratified random sampling – types of variables: qualitative and quantitative variables – continuous and discontinuous variables – scaling method S- mean – standard deviation – standard error – coefficient of variation.

UNIT III:

Coparision of means, chisquard test, student test (ANOVA ‘’portioning of variation). F test – model sums on one way ANOVA with interpretation of data – introduction to MANIVA – Statistical and their use – significance test and fixing levels of significance – use of statistical software like COSTAT and STATISTICA. Breif introduction to pie and histograms. Use of LCD.

UNIT IV:

Chromatography – priniciple, operative technique and applications of paper, TLC, adsorption chromatography, GLC and HPLC. Ion-Exchange, molecular sieve, Electrophoretic techniques – principle and technique of gel, SDS, high voltage and discontinuous electrophoresis, Isoelectric focusing, plused field gel electrophoresis and capillary electrophoresis. Spectrometry – Centrifugation techniques.

UNIT V:

X-Rays – X-Ray diffraction, crystals and detectors, quantitative analysis and applications. Radio chemical methods – Basic concepts, counting methods and applications. Autoradiography, detection and measurement of radioactivity, applications of radioisotopes in biology.

REFERENCES:

1. An introduction to practical biochemistry by David T. Plummer.
2. Laboratory Manual in Biochemistry by Pattabiraman and Acharya
3. Practical Biochemistry by J. Jayaraman.
4. Analytical Biochemistry, D. J. Homie and Hazel Peck, Longman group, 3rd edition, 1998.
5. Physical Biochemistry – Application of Biochemistry and Molecular Biology, David Friefelder, W.H Freeman and Co, 2nd Edition 1999.
6. Experimental Biochemistry, Robert Switzer and Liamgarrity, W.H. Freeman and Co, 3rd 1999.
7. Davis, G.B and C.A Parker, 1997. Writing the doctoral dissertation, Barrons Education series, 2nd edition, Pp 160, ISBN: 081208005
8. Duneary, P. 2003. Authoring a Ph. D thesis: how to plan, draft, write and finish a doctoral dissertation. Plagrave Macmillan, Pp256. ISBN 1403905843

SEMESTER II

Course Code	Course Title	L	T	P	C
23215AEC21	Enzymology	4	1	0	4

Pre-requisites Basic knowledge about catalysis, kinetics and chemical reaction mechanisms.

Course Objectives

1. Students will be introduced to the theory and practice of enzymology.
2. Mechanisms of catalysis and factors affecting catalysis will be understood
3. The kinetics of enzyme catalyzed reactions in the absence and presence of inhibitors will be studied and the options for applying enzymes and their inhibitors in medicine will be analyzed.
4. Students will learn about the applications of enzymes in research, medicine, and industry, which will prepare them for careers in industrial and biomedical research.
5. The control of metabolic pathways and cellular responses through enzyme regulation will be emphasized.

Course Outcomes **On successful completion of this course, students should be able to:**

CO1: Describe the catalytic mechanisms employed by enzymes (K1, K2 & K5)

CO2: Choose and use the appropriate methods to isolate and purify enzymes and check the purity of the enzyme .(K1,K2 , K3,K4 & K5)

CO3: Analyze enzyme kinetic data graphically, calculate kinetic parameters, determine the mechanism of inhibition by a drug/chemical and analyze options for applying enzymes and their inhibitors in

medicine (K1, K2, K3 &K4)

CO4: Explain allosterism and cooperativity and differentiate Michaelis-Menten kinetics from sigmoidal kinetics. The role played by enzymes in the regulation of vital cellular processes will be appreciated. (K1, K2 , K5, K6)

CO5: Highlight the use of enzymes in industries and biomedicine (K1,K2 & K3)

Units I

Introduction to enzymes and features of catalysis: A short history of the discovery of enzymes and how they became powerful biochemical tools. Holoenzyme, apoenzyme, cofactors, coenzyme, prosthetic groups, Classification and Nomenclature, Specificity of enzyme action-group specificity, absolute specificity, substrate specificity, stereochemical specificity. Active site, Identification of amino acids at the active site-trapping of ES complex, identification using chemical modification of amino acid side chains and by site-directed mutagenesis.

Mechanisms of enzyme catalysis: acid-base catalysis, covalent catalysis, electrostatic catalysis, metal ion catalysis, proximity and orientation effects, Low barrier H-bonds, Structural flexibility Mechanism of action of chymotrypsin

Units II

Enzyme techniques: Isolation and purification of enzymes - Importance of enzyme purification, methods of purification- choice of source , extraction, fractionation methods-based on size or mass (centrifugation, gel filtration); based on polarity (ion-exchange chromatography, electrophoresis, isoelectric focusing, hydrophobic interaction chromatography); based on solubility (change in pH, change in ionic strength); based on specific binding sites (affinity chromatography) ,choice of methods, Criteria of purity of enzymes.

Enzyme units - Katal, IU. Measurement of enzyme activity - discontinuous, continuous, coupled assays; stopped flow method and its applications. Isoenzymes and their separation by electrophoresis with special reference to LDH

Units III

Enzyme kinetics I: Thermodynamics of enzyme action, Activation energy, transition-state theory, steady-state kinetics & pre-steady-state kinetics.Single substrate enzyme catalyzed reactions -assumptions, Michaelis-Menten and Briggs-Haldane kinetics, derivation of Michaelis-Menten equation . Double reciprocal (Lineweaver-Burk) and single reciprocal (Eadie -Hofstee) linear plots, their advantages and limitations. Analysis of kinetic data- determination of Km, Vmax, kcat, and their physiological significance, Importance of kcat/Km. Enzyme inhibition: Irreversible inhibition. Reversible inhibition-Competitive, uncompetitive ,noncompetitive, mixed and substrate inhibition. Michaelis -Menten equation in the presence of competitive, uncompetitive and non-competitive inhibitors. Graphical analysis - Diagnostic plots for the determination of inhibition type. Therapeutic use of enzyme inhibitors-Aspirin, statins (irreversible inhibitors), Methotrexate (competitive inhibitor), Etoposide (non-competitive inhibitor), camptothecin (uncompetitive inhibitor).

Demonstration :Using Microsoft Excel to Plot and Analyze Kinetic Data

Units IV

Enzyme kinetics II: Allosteric enzymes: Cooperativity, MWC and KNF models of allosteric enzymes, Sigmoidal kinetics taking ATCase as an example. Regulation of amount and catalytic activity by - extracellular signal, transcription, stability of mRNA, rate of translation and degradation, compartmentation, pH, temperature, substrate concentration, allosteric effectors, covalent modification. Regulation of glycogen synthase and glycogen phosphorylase. Feedback inhibition-sequential, concerted, cumulative, enzyme-multiplicity with examples.

	Bi - Substrate reactions: Single Displacement reactions (SDR) (Ordered and Random bi bi mechanisms), Double Displacement reactions (DDR) (Ping pong mechanism), Examples, Cleland's representation of bisubstrate reactions, Graphical analysis (diagnostic plots) to differentiate SDR from DDR.
Units V	Enzyme technology: Immobilization of enzymes – methods - Reversible immobilization (Adsorption, Affinity binding), Irreversible immobilization (Covalent coupling, Entrapment and Microencapsulation, Crosslinking, Advantages and Disadvantages of each method, Properties of immobilized enzymes,. Designer enzymes-ribozymes and deoxyribozymes, abzymes, synzymes. Enzymes as therapeutic agents-therapeutic use of asparaginase and streptokinase. Application of enzymes in industry- Industrial application of rennin, lipases, lactases, invertase, pectinases, papain.
Reading List (Print and Online)	**Enzymes** | MIT OpenCourseWare | Free Online Course Materials https://ocw.mit.edu/high-school/biology/exam-prep/chemistry-of-life/enzymes/ **Enzymology** https://onlinecourses.swayam2.ac.in/cec20_bt20/preview https://mooc.es/course/enzymology/ **The active site of enzymes** https://dth.ac.in/medical/courses/biochemistry/block-1/1/index.php **Enzymes and Enzyme Kinetics** https://www.lecturio.com/medical-courses/enzymes-and-enzyme kinetics.course#/ Mechanistic enzymology in drug discovery: a fresh perspective https://www.nature.com/articles/nrd.2017.219 Enzyme Biosensors for Biomedical Applications: Strategies for Safeguarding Analytical Performances in Biological Fluids https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4934206/

Self-Study	1.Mechanistic enzymology in drug discovery 2. Enzyme Biosensors for Biomedical Applications
Recommended Texts	1.Enzymes: Biochemistry, Biotechnology and Clinical chemistry, 2nd edition, 2007, Palmer T and Bonner P; Affiliated- East West press private Ltd, New Delhi 2.Fundamentals of Enzymology, 3rd edition, 2003, Price NC and Stevens L; Oxford University Press, New York 3.Voet's Biochemistry, Adapted ed, 2011,Voet,D and Voet JG; Wiley, India 4.Lehninger Principles of Biochemistry, 8th edition, 2021, .Nelson DL and Cox MM; WH Freeman & Co, New York 5. Biochemistry, Berg JM, Stryer L, Gatto,G, 8th ed, 2015;WH Freeman & Co., New York. 6.Enzyme Kinetics and Mechanism; Cook PF, Cleland W, ;2007; Garland Science, London

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	**S**	**M**	**S**	**L**	**M**	**S**	**L**	**S**	**S**	**M**
CO 2	**S**	**S**	**S**	**S**	**M**	**M**	**L**	**S**	**S**	**S**
CO 3	**S**	**S**	**S**	**S**	**M**	**M**	**M**	**S**	**S**	**S**
CO 4	**S**	**S**	**S**	**S**	**M**	**M**	**M**	**S**	**S**	**S**
CO 5	**S**	**S**	**S**	**S**	**M**	**L**	**M**	**S**	**S**	**S**

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215AEC22	CELLULAR METABOLISM	4	1	0	4

Pre-requisites Basic knowledge on biochemical reactions such as addition, deletion, rearrangement, transfer and breaking of bonds

Course Objectives

1. Familiarize on blood glucose homeostasis
2. Provide an insight into the metabolic path way of glycogen, glycoprotein, mucopolysaccharide and peptidoglycan with clinical correlation wherever required
3. Inculcate knowledge on nucleotide metabolism and disorders associated with it
4. Provide a platform to understand the versatile role of PLP in amino acid degradation, formation of specialized products and disorders associated with ammonia detoxification
5. Educate on heme and sulphur metabolism with associated clinical manifestation

Course Outcomes **On successful completion of this course, students should be able to:**

After completion of the course, the students should be able to:

CO1. Appreciate the modes of synthesis and degradation of glucose and will be able to justify the pros and cons of maintain the blood sugar level (**K1, K2, K5**)

CO2. Gain knowledge on polysaccharide metabolism and glycogen storage disease **(K1, K2, K5)**

CO3. Acquaint with the making and braking of nucleotides **(K1,K2,K4)**

CO4. Differentiate the diverse reaction a particular amino acid can

experience **(K1,K2,K3)**

CO5. Correlate the disturbance of metabolic reactions to clinical manifestations with reference to heme and sulphur metabolism (**K1, K2, K4, K5)**

Units I Glycolysis – aerobic and anaerobic, inhibitors, and regulation. Feeder pathway- entry of hexoses into glycolysis, Galactosemia, fructosuria, Pyruvate dehydrogenase complex-mechanism and regulation. Glyoxalate cycle and its regulation. Gluconeogenesis- source, key enzymes, reaction sequence and its regulation. Blood glucose homeostasis and the role of hormones. Pentose phosphate pathway-significance and its regulation. Metabolism of glycogen and its regulation. Biosynthesis of N-linked and O-linked glycoproteins, mucopolysaccharides, Chondroitin sulphate.

Units II Oxidation of fatty acids-oxidation of saturated and unsaturated fatty acids (α, β & ω oxidation) Oxidation of fatty acids with odd and even numbered carbon atoms. Regulation of β oxidation. Ketogenesis and its regulation. Biosynthesis of fatty acid–saturated and unsaturated, chain elongation, regulation. Biosynthesis of prostaglandins, thromboxanes and leukotrienes and hydroxyl eicosanoic acids. Biosynthesis and degradation of triacylglycerol, phosphoglycero lipids-lecithin, cephalin, plasmalogens and phosphatidyl inositol, Sphingolipid-sphingomyelin, cerebrosides, sulfatides, and gangliosides. Cholesterol biosynthesis and its regulation. Lipoprotein metabolism-chylomicrons, VLDL, HDL and LDL.

Units III Metabolism of nucleotides- *De novo* synthesis and salvage pathways of purine and pyrimidine nucleotides. Regulation and inhibitors of nucleotide biosynthesis. Role of ribonucleotide reductase and its regulation. Degradation of purine and pyrimidine nucleotides.

Units IV	Biosynthesis of non- essential amino acids.- Role and biological significance of glutamate dehydrogenase, glutamine and asparagine synthetase, lysine, proline and phenylalanine hydroxylase. Interconversion of amino acids - proline to glutamate, methionine to cysteine, serine to glycine. Biosynthesis of spermine and spermidine. Degradation of amino acids –glucogenic and ketogenic amino acids. Formation of acetate from leucine and aromatic amino acid, pyruvate from cysteine, threonine and hydroxy proline, α-keto glutarate from histidine and proline, succinate from methionine, threonine, valine and isoleucine, Oxaloacetate from aspartate, glycine and serine.
Units V	Biosynthesis and degradation of heme. Jaundice-classification, pathology and Differential diagnosis Oxidation and reduction of inorganic sulphur compounds by microbes and plants. Sulpho transferases and their biological role-rhodanases, sulphatases , 3-mercapto pyruvate sulphur transferases. Mucopolysaccharidoses - Hunter syndrome, Sanfilippo syndrome and Maroteaux-Lamy syndrome. Oxidation of cysteine to sulphate and inter conversion of sulphur compounds.
Reading List (Print and Online)	1. https://www.embopress.org/doi/full/10.1038/msb.2013.19 2. https://people.wou.edu/~guralnl/450Glycogen%20metabolism.pdf 3. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3243375/ 4. https://www.researchgate.net/publication/334458898_Urea_Cycle 5.https://www.researchgate.net/publication/51233381_Heme_biosynthesis_and_its_regulation_Towards_understanding_and_improvement_of_heme_biosynthesis_in_filamentous_fungi 6.https://www.researchgate.net/publication/349746691_Microbial_Sulfur_Metabolism_and_Environmental_Implications
Self-study	1. Cori’s Cycle and Glucose- Alanine Cycle 2. Coenzymes involved in Methanogenesis
Books Recommended	1.David L.Nelson and Michael M.Cox (2012) Lehninger Principles of Biochemistry (6th ed), W.H.Freeman

2.Voet.D and Voet. J.G (2010) Biochemistry , (4th ed), John Wiley & Sons, Inc.
3.Metzler D.E (2003). The chemical reactions of living cells (2nd ed), Academic Press.
4. Zubay G.L (1999) Biochemistry , (4th ed), Mc Grew-Hill.
5. Textbook of Biochemistry with Clinical Correlations, 7th Edition,Thomas M. Devlin (Editor), Wiley
6. Human Biochemistry – James M.Orten & Otto.W.Neuhan- 10th edn- The C.V.Mosby Company

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	M	S	M	S	M	S	S	S	M
CO 2	S	M	S	S	S	M	S	S	S	M
CO 3	S	M	S	S	S	M	S	S	S	S
CO 4	S	M	S	M	S	M	S	S	S	M
CO 5	S	M	S	S	S	M	S	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215AEC23	CLINICAL BIOCHEMISTRY	4	1	0	4

Pre-requisites, if any: The student should have a basic knowledge of body fluids and their composition and metabolism; anatomy and physiology of vital organs.

Course Objectives

1. To understand the need and methods of various biological sample collection.
2. To explicitly understand the etiopathogenesis, symptoms and complications of metabolic and hormonal disorders and the relevant diagnostic markers
3. To emphasize the diagnostic significance of serum enzymes in different pathologies and other Laboratory investigations of diagnostic importance so as to differentiate normal from disease
4. To conceive the role of inherited genes in inborn errors of metabolism and methodologies pertaining to *in utero* diagnosis and post-natal screening.
5. To get updated about electrolyte and hormonal imbalances and the biochemical tests to diagnose them.

Course Outcomes

CO1. To appreciate the biological significance of sample collection and awareness of the diagnostic/screening tests to detect common non-communicable diseases so as to understand role of laboratory investigations for biochemical parameters and understand the disorders associated with blood cells

CO2. To understand the etiology of metabolic diseases like diabetes and atherosclerosis and avoid such lifestyle disorders by healthy eating and correlate the symptoms with underlying pathology based on diagnostic and prognostic markers.

CO3. To understand the diagnostic application of serum/plasma enzymes to correlate their levels with the organ pathologies associated with specific diseases.

CO4. To appreciate the role of pre and post-natal diagnosis leading to healthy progeny.

CO5. To link the serum hormone levels and clinical symptoms with underlying hormonal disturbances. To review the onward transmission of signal via downstream signaling molecules from cell surface to the nucleus by different pathways by comparing and contrasting them and critically evaluate the network between them resulting in the biological outcome.

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Unit I

Biochemical investigations in diagnosis, prognosis, monitoring, screening: Specimen collection – blood, (primary /Secondary specimen)., urine and CSF. Preservation of biological specimens -blood, urine, CSF and amniotic fluid. ; . Biological reference ranges;

Disorders of blood cells: Hemolytic, iron deficiency and aplasticanemia and diagnosis, sickle cell anaemia, thalassemia HBA1C variants. Porphyrias, Thrombocytopenia, Causes of leucopenia, leukemia and leucocytosis.

Disorders of blood clotting mechanism - Von willebrand's disease, Hemophilia A, B and C, diagnostic test for clotting disorders,
D-dimer and its clinical significance

Unit II

Diabetes mellitus: pathology and complications: Acute changes; Chronic complications: Diabetic nephropathy, neuropathy, retinopathy and Diabetic foot ulcers, Random/Fasting/PP glucose testing, Impaired glucose tolerance (IGT), Impaired fasting glucose (IFT), Diagnosis-by GTT, Pre-diabetes, Gestational DM ,Glycosylated Haemoglobin (HBA1c) ; Glycated albumin., Hypoglycaemia and critical alert value for glucose. Markers of complications of Diabetes mellitus: Metabolic syndrome, Lipid profile &lipoproteinemia, Atherosclerosis, Diabetic nephropathy, Micralbuminuira, eGFR.
Point of care testing for glucose (Glucometers) and continuous glucose monitoring (CGM) : principle and its use. Major groups of anti-diabetic drugs. Diet and life style modifications

Unit III

Diagnostic Enzymology: Clinically Important Enzymes and Isoezyme as diagnostic markers: Clinical significance of AST, ALT, ALP, ACP, CK, γ-GT, amylase, pseudocholinesterase and their pattern in .Myocardial infarction; Liver disease, Bone disease, Muscle disease, Cancer (tumor markers), GI tract pancreatitis); Enzymes as therapeutic agents.

Pre- and post-natal testing: Amniocentesis, prenatal detection of inborn errors of metabolism in developing fetus- Autosomal recessive mode of inheritance-cystic fibrosis, X linked recessive inheritance-Duchenne muscular dystrophy. New born screening (NBS) for In born errors of metabolism, Tandem mass spectrometry application in NBS.

Unit IV **Liver function tests:** Liver function test panel, Fatty liver . Plasma protein changes in liver diseases. Hepatitis A ,B and C. Cirrhosis and fibrosis. Portal hypertension and hepatic coma.Acute phase proteins -CRP, Haptoglobins, α-fetoprotein, ferritin and transferrin and their clinical significance, Interpreting serum protein electrophoresis. Inflammatory markers (cytokines such as TNF-alpha IL6 and others)

Unit V **Renal function tests** - tests for glomerular and tubular function-Acute and chronic renal failure-Glomerulonephritis, Nephrotic syndrome, uraemia-urinary calculi-Nephrocalcinosis and Nephrolithiasis-causes, pathology and symptoms. Chronic kidney disease. Dialysis-Hemodialysis and peritoneal dialysis.

Electrolyte disorder : calcium: hypercalcemia and hypocalcemia; Calcium homoestasis in Blood; phosphate: hyperphosphatemia or hypophosphatemia; Clinical significance: Potassium: hyperkalaemia and hypokalaemia, Sodium: hypernatremia and hyponatremia; Chloride: hyperchloremia, hyporchloremia

Hormonal disorders and diagnostics: T3, T4 and TSH in the diagnosis of thyroid disorders; Diagnostic methods for disorders associated with adrenal, pituitary and sex hormones - Addison's disease, Cushing's syndrome, pituitary tumour, Hypopituitarism, Hypogonadism

Reading List (Print and Online)

1. **Utility of HIL in Clinical Chemistry:** https://www.aacc.org/science-and-research/clinical-chemistry-trainee-council/trainee-council-in-english/pearls-of-laboratory-medicine/2018/utility-of-hil-in-clinical-chemistry

2. **Pre, Post and Analytical Errors in Clinical Chemistry laboratory**
DOI: 10.7860/NJLM/2016/22587:2173
https://doi.org/10.2147/JMDH.S286679

3. **Standards of Medical Care in Diabetes—2022 Abridged for Primary Care Providers**
https://diabetesjournals.org/clinical/article/40/1/10/139035/Standards-of-Medical-Care-in-Diabetes-2022

https://doi.org/10.2337/diaspect.16.1.32

http://www.ngsp.org/

4. Quality control in clinical laboratory

https://www.researchgate.net/publication/335830829_Quality_Control_in_a_Clinical_Laboratory

https://labpedia.net/quality-control-of-the-clinical-laboratory/

https://journals.sagepub.com/doi/full/10.1016/j.jala.2008.12.001

https://doi.org/10.1016/B978-0-12-407821-5.00004-8

https://www.westgard.com/clia.htm

https://www.labroots.com/webinar/bio-rad-unity-solution-molecular-quality-control-data-management

Self-Study

1. **Potential sources of variability in the estimation of the analytes:**

   Pre-analytical phase: acceptance rejection criteria in terms of haemolysis/icteric/lipemia (HIL) interferences

   Analytical phase: Linearity, detection limits precision, accuracy, specificity, sensitivity; Total Allowable Error. (Definitions and examples).

   Post-analytical phase **:** Units of reporting of clinical chemistry parameters-

2. **Interpretation of results in clinical chemistry based on laboratory investigations and quality control:**

- critical / alert values
- American Diabetes Association (ADA) Standards of Medical Care in Diabetes (yearly update); HBA1C testing :NGSP
- Case studies to review
- Quality control for clinical chemistry in laboratory

Recommended Texts

1. ThomasM.Devlin (2014) Textbook of Biochemistry with Clinical Correlations (7th ed). John Wiley & Sons
2. Montgomery R, Conway TW, Spector AA (1996),Biochemistry: A Case-Oriented Approach (6th ed), Mosby Publishers, USA.
3. Tietz Fundamentals of Clinical Chemistry and Molecular Diagnostics

(2018) (8th ed),Saunders

4. Dinesh Puri, (2020) Text book of Biochemistry: A clinically oriented approach – 4th Edition, Elsevier.
5. 5. M.N.Chatterjee and Rana Shinde (2012).Textbook of Medical Biochemistry (8th ed), Jaypee Brothers Medical Publishers.
6. Clinical Case Discussion In Biochemistry A Book On Early Clinical Exposure (ECE), Poonam Agrawal , 2021, CBS Publishers & distributors pvt. Ltd

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Observe, Explain.

Analyse (K4)- Finish procedure in stepwise manner, Differentiation between various ideas, Map knowledge

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion, Debating, Presentation

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	S	M	S	S	S	S	M	M	S
CO 2	S	M	S	M	S	S	S	M	M	M
CO 3	S	S	S	S	S	M	S	S	M	M

CO 4	**S**	**M**	**M**	**M**	**S**	**M**	**S**	**S**	**S**	**M**
CO 5	**S**	**M**	**S**	**M**	**S**	**S**	**S**	**S**	**S**	**S**

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215SEC24L	Enzymology , Microbiology and Cell Biology Lab	0	0	4	4

Pre-requisites Knowledge on basic principles, Instrumentation of Biochemical techniques and metabolic reactions

Course Objectives

1. To inculcate skill in students enabling them to apprehend the wider knowledge about principles and techniques to be employed for the assayof enzymes under investigation.
2. To inculcate the knowledge of isolation and purification techniques ofenzymes using alkaline phosphatase as an example
3. To perform experiments to study the factors affecting enzyme activity
4. To achieve training in assay of enzymes
5. To achieve training in basic microbiological techniques – preparation ofculture, sterilization and staining methods.
6. To perform the blood grouping test and to prepare blood smear to studydifferent types of blood cells
7. To learn molecular biology techniques like Gel electrophoresis and Blotting techniques
8. To introduce industrial visit so that students may be aware of actual need of the industry and various opportunities available

Course Outcomes **On successful completion of this course, students should be able to:**

After completion of the course, the students should be able to:

CO1. The student will be able to employ the relevant techniques for isolation and purification of enzymes and gain skill in kinetic studies which is essential for research activity (K1,K2, K4)

CO2. Student will acquire ability in performing enzyme assay, and explicate the methods that form the basis of enzyme characterization. (K1,K2,K4)

CO3. Learn the Basic concepts in microbiology and cell biology which will be helpful for interdisciplinary research work. (K1,K3,K4)

CO4. Students will be trained in separation techniques used in molecular Biology which will be supportive in their future research (K1,K3,K4 & K6)

CO5. Industrial visits will provide the students with an opportunity to learn practically through interaction, working methods and employment practices. Students will have an exposure to Industrial standard and current work practices (K1,K2,K3,K4 & K6)

Units I Enzymology

Alkaline Phosphatase

a. Isolation of Alkaline Phophatase from goat kidney.
b. Purification of alkaline phosphatase
c. Checking the purity using SDS-PAGE
d. Determination of optimum pH and temperature of alkaline phosphatase.
e. Determination of specific activity and Km of alkaline phosphatase.
f. Effect of activators and inhibitors on the activity of alkaline phosphatase.

	Assay of enzymes a. Salivary Amylase b. Acid Phosphatase
Units II	Microbiology a. Safety measures and Good Laboratory Practices in microbiology laboratory b. Sterilization,Culture and inoculum preparation c. Staining of bacteria – Gram Staining
Units III	Physiology & Cell Biology a. Test for blood grouping (Haemagglutination). b. Peripheral Blood smear –Staining and Interpretation
Units IV	Group Experiments a. Separation of proteins based on molecular weight by SDS PAGE b. Agarose gel electrophoresis of genomic DNA
Units V	Industrial visit can be organised to students through Academia –Industry collaborative Program
Reading List (Print and Online)	1.https://www.researchgate.net/publication/337146254_Kinetic_studies_with_alkaline_phosphatase 2.https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4846332/ 3.https://www.ijsr.net/archive/v3i8/MDIwMTU0MDk=.pdf 4.https://www.researchgate.net/publication/349318898_ABC_of_Periheral_smear 5.https://ncdc.gov.in/WriteReadData/l892s/File608.pdf 6.https://www.ncbi.nlm.nih.gov/books/NBK562156/
Self-Study	1. Preparation of Buffers and pH measurement 2. Michaelis-Menten equation and Lineweaver Burk plot
Books Recommended	1. David Plummer (2001) An Introduction to Practical Biochemistry (3rd ed) McGraw Hill Education (India) Private Ltd 2. Jayaraman, J (2011), laboratory Manual in Biochemistry, New age publishers 3. Fundamentals of Enzymology; 3rd Edn. Nicholas C. Price and Lewis

Stevens, Oxford University Press (2012).

4. Enzymes: A Practical Introduction to Structure, Mechanism, and Data Analysis; Robert A. Copeland , Wiley-VCH Publishers (2000).
5. Cappuccino JG & Sherman N (2005). Microbiology-A Laboratory Manual, Pearson Education Inc
6. Practical Enzymology, Second Revised Edtion: Hans Bisswanger, Wiley – Blackwell; 2 edition (2011)

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	**PO 2**	**PO 3**	**PO 4**	**PO 5**	**PO 6**	**PO 7**	**PO 8**	**PO 9**	**PO 10**
CO 1	**S**	**S**	**S**	**S**	**M**	**S**	**L**	**S**	**M**	**S**
CO 2	**S**	**S**	**S**	**S**	**M**	**S**	**L**	**S**	**M**	**S**
CO 3	**S**	**S**	**S**	**S**	**M**	**S**	**M**	**S**	**M**	**S**
CO 4	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**
CO 5	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC25A	Energy and drug metabolism	4	1	0	3

Pre-requisites Basic knowledge on biochemical reactions such as addition, deletion, rearrangement, transfer and breaking of bonds

Course Objectives

1. Familiarize on concepts of enthalpy, entropy, free energy, redox system, biological oxidation and high energy compounds
2. Provide an insight into the relationship between electron flow and phosphorylation
3. Inculcate knowledge on processes involved in converting light energy to chemical energy and associated food production by autotrophs
4. Provide a platform to understand the versatile role of Krebs cycle, transport of NADH across mitochondrial membrane and energetics
5. Educate on the various phases xenobiotic metabolism

Course Outcomes **On successful completion of this course, students should be able to:**

After completion of the course, the students should be able to:

CO1. Appreciate the relationship between free energy and redox potential and will be able to justify the role of biological oxidation and energy rich compounds in maintaining the energy level of the system (**K1,K2,K3,K4**)

CO2. Gain knowledge on role of mitochondria in the production of energy currency of the cell **(K1, K2, K5, K6)**

CO3. Acquaint with the process of photosynthesis **(K1,K2,K5)**

CO4. Comprehend on the diverse role of TCA cycle and the energy obtained on complete oxidation of glucose and fatty acid

(K1,K2,K4,K5)

CO5. Correlate the avenues available to metabolize the xenobiotics (**K1, K2,K4,K5)**

Units I Thermodynamic- principles in biology- Concept of entropy, enthalpy and free energy change. Redox systems. Redox potential and calculation of free energy. Biological oxidation – Oxidases, dehydrogenases, hydroperoxidases, oxygenases. Energy rich compounds – phosphorylated and non-phosphorylated. High energy linkages.

Units II Electron transport chain-various complexes of ETC, Q-cycle. Inhibitors of ETC. Oxidative phosphorylation-P/O ratio, chemiosmotic theory. Mechanism of ATP synthesis - role of F0-F1 ATPase, ATP-ADP cycle. Inhibitors of oxidative phosphorylation ionophores, protonophores .Regulation of oxidative phosphorylation

Units III Light reaction-Hills reaction, absorption of light, photochemical event. Photo ETC-cyclic and non-cyclic electron flow. Photophosphorylation-role of CF0-CF1 ATPase. Dark reaction- Calvin cycle, control of C3 pathway, and Hatch-Slack pathway (C4 pathway), Photorespiration. Synthesis and degradation of starch

Units IV Interconversion of major food stuffs. Energy sources of brain, muscle, liver, kidney and adipose tissue. Amphibolic nature of Citric acid cycle. Anaplerotic reaction. Krebs cycle, Inhibitors and regulation of TCA cycle. Transport of extra mitochondrial NADH – Glycerophosphate shuttle, malate aspartate shuttle. Energetics of metabolic pathways – glycolysis, (aerobic and anaerobic) ,citric acid cycle, beta oxidation

Units V Activation of sulphate ions – PAPS, APS, SAM and their biological role. Metabolism of xenobiotics – Phase I reactions – hydroxylation, oxidation and reduction. Phase II reactions – glucuronidation, sulphation, glutathione conjugation, acetylation and methylation. Mode of action and factors affecting the activities of xenobiotic enzymes.

Reading List (Print and Online)

1.https://chemed.chem.purdue.edu/genchem/topicreview/bp/ch21/gibbs.php

2.https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7767752/#:~:text=The%20mitochondrial%20electron%20transport%20chain,cellular%20ATP%20through%20oxidative%20phosphorylation.

3. https://www.researchgate.net/figure/Oxidative-phosphorylation-in-mitochondrial-electron-transport-chain-ETC-and-proton_fig1_230798915

4.https://www.lyndhurstschools.net/userfiles/84/Classes/851/photosynthesis%20light%20&%20dark%20reactions%20ppt.pdf?id=560837

5.https://bajan.files.wordpress.com/2010/05/amphibolic-nature-of-krebs-cycle.pdf

6.https://www.sciencedirect.com/topics/medicine-and-dentistry/xenobiotic-metabolism#:~:text=Xenobiotic%20metabolism%20can%20be%20defined,more%20readily%20excreted%20hydrophilic%20metabolites

Self-Study

1. Calculation of Keq and $\triangle$ G
2. Interrelationship of carbohydrate, protein, and fat metabolism-role of acetyl CoA

Recommended Texts

1.David L.Nelson and Michael M.Cox (2012) Lehninger Principles of Biochemistry (6th ed), W.H.Freeman

2. Robert K. Murray, Darryl K. Granner, Peter A. Mayes, and Victor W. Rodwell (2012), Harper's Illustrated Biochemistry, (29th ed), McGraw-Hill Medical

3. Metzler D.E (2003). The chemical reactions of living cells (2nd ed), Academic Press.

4. Zubay G.L (1999) Biochemistry , (4th ed), Mc Grew-Hill.

5. Devlin RM (1983) Plant Physiology (4th ed), PWS publishers

6.Taiz L , Zeiger E (2010), Plant Physiology (5th ed), Sinauer Associates, Inc

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	**PO 2**	**PO 3**	**PO 4**	**PO 5**	**PO 6**	**PO 7**	**PO 8**	**PO 9**	**PO 10**
CO 1	**S**	**S**	**S**	**M**	**S**	**M**	**S**	**S**	**S**	**M**
CO 2	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**
CO 3	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**
CO 4	**S**	**M**	**S**	**M**	**S**	**M**	**S**	**S**	**S**	**L**
CO 5	**S**	**M**	**S**	**S**	**S**	**M**	**S**	**S**	**S**	**S**

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC25B	**Neuro Biochemistry**	4	1	0	3

Aim:

The course aims to provide students with a basic understanding of:

- the principles and major mechanisms of metabolic control and of molecular signalling by hormones;
- the control of cell proliferation

Course Objective:

By the end of the course, students should be able to:

- demonstrate knowledge and understanding of the molecular machinery of living cells;
- demonstrate knowledge and understanding of the principles that govern the structures of macromolecules and their participation in molecular recognition;
- use basic laboratory skills and apparatus to obtain reproducible data from biochemical experiments.

Course outcomes:

- CO1 To understand various neurological system
- CO2 Recognize the need for, and engage in life-long learning in neurological system
- CO3 To understand various Exocytosis of neurotransmitter
- CO4 To able to understand DNA microarrays, Methodology, types and applications
- CO5 To acquire knowledge related to LEARNING AND MEMORY
- CO6 Gain knowledge of contemporary issues
- CO7 to understand biochemistry of vision and muscle contraction

UNIT I : NERVOUS SYSTEM

Structure and function of the brain. Central Nervous System, Peripheral and Autonomic Nervous system. Cells of Nervous System – Neurons, Astrocytes, Glial cells, Oligodendrocytes and Schwan cells. Chemical composition of brain – utilization and uptake of glucose and amino acids, Blood – Brain barrier.

UNIT II: NEUROTRANSMISSION

Membrane potentials, Resting potential – Depolarization, repolarization and hyperpolarization, Action potential. Mechanism of axonal neurotransmission. Membrane channels – Types of channels, ion gated, voltage gated, chemically gated, mechanically gated and responsive to intracellular messengers. DISEASES OF NERVOUS SYSTEM Molecular basis of Parkinson's disease, Alzheimer's disease, Schizophrenia, Myasthenia gravis and Multiple sclerosis.

UNIT III: NEUROTRANSMITTERS

Synthesis, storage, release, uptake, degradation and action of neurotransmitters. Acetyl choline, GABA, Serotonin, Dopamine, Glutamate, Aspartate, Nitrous oxide, etc. Neuropeptides. Synaptic transmission – Cholinergic receptors – Nicotinic and Muscarinic receptors, Agonists and Antagonists – their mode of action and effects. Adrenergic receptors, serpentine receptors and intracellular signaling. Fast and slow receptors. Exocytosis of neurotransmitter – Role of synapsins, synaptogamins, SNAP, SNARE and other proteins in docking, exocyotosis and recycling of vesicles.

UNIT IV : LEARNING AND MEMORY

Mechanism of short term memory and Long Term Potentiation. NMDA and AMPA glutamate receptors. Retrograde messengers in synaptic transmission. Role of CAM kinase II, Calcium, protein kinases, cAMP, NO, Calpain and other proteins in memory and learning process. Synaptic plasticity INTERACTION OF DRUGS WITH CNS Mechanism of action of anesthetics, analgesics, hallucinogens, depressants, stimulants and toxins on the nervous system. Addiction and drugs of abuse.

UNIT V: BIOCHEMISTRY OF VISION AND MUSCLE CONTRACTION:

Rod and cone cells, visual cycle, mechanism and regulation of vision, color vision. Thick and thin filaments, interaction of actin and myosin muscle contraction, role of calcium and regulation of muscle contraction. Smooth muscle contraction and its regulation

REFERENCE

1. Neurochemistry by Ferdinand Hucho, VCH Publication, 1986

2. Molecular cell Biology,by Lodish, Baltimore, et al W.H. Freeman & Co. !996

3. Basic Neurochemistry by M. P. Spiegel

Course Code	Course Title	L	T	P	C
23215SEC26	Nutritional Biochemistry	4	0	0	3

Pre-requisites, if any: **BASIC KNOWLEDGE ON FOOD , NUTRITION & DIETETICS, AND METABOLISM OF NUTRIENTS.**

Course Objectives

1. To understand basic concepts involved in growth , health, nutrition, physiology and metabolism
2. To discuss the concepts and applications of nutrition in correlation with biochemistry
3. To define nutritional needs in healthy individuals and modification of diet during illness.

Course Outcomes

After completion of the course, the students should be able to:

CO1. Plan a balanced diet based on an individual's energy requirement, Assess nutritional status of an individual(K3, K4, K5)

CO2. Describe the biochemical , physiological and nutritional functions of macronutrients and their integrated role. Understand the role played by antinutritional factors(k! to K6)

CO3. Evaluate the functions of vitamins and minerals ,and fluids and electrolyte balance in different physiological states and in sports persons(K1 to K6)

CO4. Identify nutritional deficiency conditions , its prevention and dietary management((K3,K4)

CO5. Acquire knowledge about the importance of balanced diet and diet therapy (k5,K6)

Units I

Basic concepts - Nutrition - Food groups and balanced diet. Novel Foods. Calorific value of foods: Direct and indirect calorimetry.

Empty calories. Basal metabolic rate: Factors affecting BMR. SDA and physical activity. Calculation of day's energy requirement. Assessment of nutritional status. Lactose intolerance. Nutritional requirement and biochemical changes in different physiological states -infancy, childhood, pregnancy, lactation, and ageing. Sports nutrition.

Units II Elements of nutrition - Plant and animal sources of simple and complex carbohydrates, fats and proteins and their requirement. Biological significance, deficiency and toxicity of macronutrients and micronutrients. Role of dietary fibre. Protein sparing action of carbohydrates and fats. Essential amino acids. Essential fatty acids. Effects of naturally occurring food toxins, preservatives, additives, alcohol and tobacco on health.

Units III Vitamins and Minerals- Dietary sources, classification, biochemical functions, requirements, absorption, metabolism and excretion. Vitamin B complex as coenzyme. Nutritional significance of dietary calcium, phosphorus, magnesium, iron, iodine, zinc and copper.

Units IV Malnutrition - Diseases arising due to Protein - Calorie Malnutrition and undernutrition (Kwashiorkor and Marasmus), Prevention of malnutrition. Deficiency diseases associated with vitamin B complex, vitamin C and A, D, E & K vitamins - Mineral deficiency diseases - aetiology, sign and symptoms and dietary supplementation. Enrichment and fortification (vitamins and minerals)

Units V Nutrition in diseases - Aetiology, signs and symptoms , treatment and dietary management during fever(Typhoid and Malaria) and infectious diseases(COVID-19), Jaundice, hyper acidity (Ulcer), Atherosclerosis, Hypertension, kidney diseases and diabetes in adults. Starvation and Obesity. Inter-relationship of nutrition, infection,

immunity and poverty

Reading List (Print and Online)

1. https://www.jmedscindmc.com/article.asp?issn=1011-4564;year=2014;volume=34;issue=5;spage=211;epage=213;aulast=Shrivastava
2. https://www.researchgate.net/figure/Relationship-between-malnutrition-infection-and-immunity-Malnutrition-is-considered-the_fig1_280722727
3. https://en.wikipedia.org/wiki/Novel_food
4. https://www.chemicalsafetyfacts.org/preservatives/
5. https://www.sciencedirect.com/topics/agricultural-and-biological-sciences/food-enrichment

Self-Study

1. Antabuse drugs and food
2. Selection of foods and market visit, reading and understanding the food labels

Recommended Texts

1. Srilakshmi. E .(2016) Nutrition Science, New Age International Publishers.
2. Mahan, Kathleen L. (2004) Krause's Food, Nutrition and Diet Therapy, W.B.Saunder's 11th Edition
3. Andreas M. Papas (1998). Antioxidant Status, Diet, Nutrition, and Health (1st ed) CRC Press.
4. M. Swaminathan (1995) Principles of Nutrition and Dietetics. Bappco
5. Margaret Mc Williams (2012). Food Fundamentals (10th ed) Prentice Hall
6. Tom Brody (1998) Nutritional Biochemistry (2nd ed). Academic Press, USA

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create(K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	**PO 2**	**PO 3**	**PO 4**	**PO 5**	**PO 6**	**PO 7**	**PO 8**	**PO 9**	**PO 10**
CO 1	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**M**	**M**
CO 2	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**M**	**M**
CO 3	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**M**	**M**
CO 4	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**M**	**L**
CO 5	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**M**	**M**

S-Strong M-Medium L-Low

SEMESTER III

Course Code	Course Title	L	T	P	C
23215AEC31	Industrial Microbiology	5	1	0	4

Pre-requisites, if any: Basic Knowledge of Microbiology and microbial techniques

Course Objectives

To gain knowledge of the structure, classification and use of microorganisms in various industries.

To know various fermenter designs, culture systems and the application of fermentation process in industry.

To understand the production and purification of fermented products and their industrial applications.

Understand the basic concepts of food and agricultural microbiology.

Course Outcomes

CO1.Students will be able to understand the structure and classification of microorganisms (K2 , K4)

CO2.Gain knowledge of the uses of microorganisms in various industrial applications (K3 , K4)

CO3.Understand the concepts of fermentation process, harvest and recovery. (K1 , K5)

CO4.Students will know the types of microbial fermentation processes and their applications in pharmaceutical industry. (K2 , K3)

CO5.Students will learn about the use of microorganisms in beverages, diary and food industries. (K3 , K6)

Units I

Structure of bacteria, fungi and viruses and their classification. Types and characteristics of microorganisms used in Industry (a) Food Industry (b) Chemical Industry (c) Pharmaceutical Industry

Units II

Fundamentals and principles of microbial fermentation techniques – application in industry and pharmaceutical Biochemistry. Fermentation – types, techniques, design and operation of fermenters including addition of

	medium. Types and characteristics of microorganisms, environmental conditions required for the growth and metabolism of industrially and pharmaceutically important microbes. Sterilization methods in fermentation techniques, air, gas, culture medium sterilization. Steam-filtration and chemicals. Types and constituents of fermentative culture medium and conditions of fermentations, Antifoaming devices.
Units III	Recovery and estimation of products of fermentation- Production of ethanol, acetic acid, glycerol, acetone, butanol and citric acid by fermentation. Production of Enzymes- amylase, protease, lipase,Production of pharmaceuticals by fermentation– penicillin, streptomycin, tetracycline, riboflavin, vitamin B12.Beverages-wine, beer and malt beverages.
Units IV	Food Microbiology: Production of dairy products-bread, cheese and yoghurt (preparation and their types). Food borne diseases- Bacterial and Non- Bacterial. Food preservation - Principles–Physical methods: temperature (low, high, canning, drying), irradiation, hydrostatic pressure, high voltage pulse, microwave processing and aseptic packaging, Chemical methods - salt, sugar, organic acids, SO_2, nitrite and nitrates, ethylene oxide, antibiotics and bacteriocins.
Units V	Agricultural Microbiology: General Properties of soil, microorganisms in soil – decomposition of organic matter in soil. Biogeochemical cycles, nitrogen fixation, Production of bio fertilizers and its field applications – Rhizobium, azotobacter, blue green algae, mycorrhizae, azospirilium, Production of biofuels (biogas- methane), soil inoculants.
Self-Study	Micro-organisms in food processing and pharma industries Upstream and Downstream processes in Biopharma
Reading List (Print and Online)	Industrial biotechnology: https://nptel.ac.in/courses/102/105/102105058/ Bioreactors: https://nptel.ac.in/courses/102/106/102106053/ Food Microbiology: https://nptel.ac.in/courses/126/103/126103017/ Agriculture Microbiology:

https://www.youtube.com/watch?v=f7UXyVImZ_c

Recommended Texts

Food Microbiology: An Introduction: 4th edition,Matthews KR, Kniel KE, Montville TJ; American Society for Microbiology

Food, Fermentation and Micro-Organisms,2nd edition, Charles, BW; Blackwell Science Ltd

Microbiology. 5th edition , Pelczar MJ, Chan ECS and Krieg NR; McGraw Hill Book Company.

Text book of Microbiology:11th edition, Ananthanarayanan R and Paniker CKJ; Universities Press (India) Pvt. Ltd.

Food Microbiology, 3rd edition, Frazier WC and Westhoff DC;Tata McGrawHill Publishing Company Ltd, New Delhi

New Methods of Food Preservation:1st edition, Gould GW; Springer Manual of Industrial Microbiology and Biotechnology: 3rd edition, Baltz

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	**PO 2**	**PO 3**	**PO 4**	**PO 5**	**PO 6**	**PO 7**	**PO 8**	**PO 9**	**PO 10**

CO 1	S	S	M	S	S	S	M	M	S	S
CO 2	S	M	S	S	M	S	S	M	M	M
CO 3	S	M	L	S	M	M	S	S	M	S
CO 4	M	S	S	S	L	M	S	M	S	M
CO 5	S	S	M	S	S	M	M	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215AEC32	Molecular Biology	4	1	0	4

Pre-requisites, if any: Knowledge of the basics of genetics, cell biology and molecular biology.

Course Objectives

1. To introduce the students to the process of inheritance, concepts of genes, genome, chromatin and chromosomes.
2. To impart a thorough understanding of the key events of molecular biology, including the mechanisms of DNA replication, transcription and translation along with DNA repair mechanisms.
3. To provide a detailed understanding of post transcriptional and posttranslational modifications and processing of eukaryotic RNA and proteins
4. To give a detailed explanation of transcriptional regulation with lac operon and tryptophan operon as examples
5. To impart adequate information of the types of regulatory RNAs along with key concepts of gene silencing

Course Outcomes After completion of the course, the students should be able to:

CO1: Comprehend the organization of genomes, the molecular basis of DNA replication, recombination and transposition, the significance of these processes, the various ways in which the DNA can be damaged leading to mutations and lesions and the different ways in which they are repaired.(K1,K2,K3,K5)

CO2: Gain knowledge about how genes are transcribed and translated in prokaryotes and eukaryotes and how these processes are regulated, recognize the nature of the genetic code and the various experimental approaches used to crack the code (K1,K2,K3,K4,K5)

CO3: Acquire knowledge of the molecular basis of RNA processing and

RNA splicing and the various human pathologies that can result from defects of RNA modification. (K1,K2,K4,K5)

CO4: Comprehend the techniques of gene silencing and its applications.(K1,K2,K3,K4,K5,K6)

CO5: Apply the knowledge they have gained in understanding the above vital life processes to enhancing their analytical and problem-solving skills and develop an interest to pursue high quality research. (K2,K3,K4,K5,K6)

Units I

Mendel's laws of inheritance-dominance-complete, incomplete and co-dominance, multiple alleles-gene mapping in haploids and diploids, recombination mapping- restriction mapping- modes of gene information transfer in bacterial- conjugation, transformation and transduction. The bacterial chromosome, the eukaryotic genome- chromosome structure – Histones, Nucleosome, chromatin- heterochromatin, euchromatin, chromatin remodeling, DNAase hypersensitive sites, genome organization – the C-value paradox, reassociation kinetics, repetitive sequences, gene amplification, telomeres, pseudogenes, split genes, organelle genomes – mitochondrial and chloroplast genome.

Units II

DNA replication and repair: Enzymes of replication, prokaryotic replication mechanisms, primosome & replisomes, eukaryotic DNA replication, the role of topoisomerases and telomerase, regulation of replication, difference between prokaryotic and eukaryotic replication. Mutations -Types of mutations, mechanisms of mutations, mutagenic agents. DNA repair mechanisms – Direct repair, excision repair, mismatch repair, recombination repair, SOS response, eukaryotic repair systems. Recombination and mobile genetic elements- the Holliday model, the general recombination in *E.coli*, site specific recombination,

transposons and retroposons.

Units III

Transcription – Prokaryotic transcription-subunits of RNA polymerase, E. coli promoters, sigma factor and promoter recognition, alternative sigma factors, initiation, elongation, Rho-dependent and independent termination of transcription. Eukaryotic transcription- Initiation, promoter elements, RNA polymerases, transcription factors, regulatory sequences in eukaryotic protein – coding genes, CpG islands, enhancers.

Translation – organization of the ribosome, the genetic code, evidence for a triplet code, deciphering the genetic code, wobble hypothesis, deviation in the genetic code, unusual codons. activation, initiation, elongation and termination of translation in E. coli. The role of tRNA and rRNA, suppressor tRNAs and inhibitors of protein synthesis., Comparison of prokaryotic translation with eukaryotic translation.

Units IV

Regulation of gene expression in prokaryotes-– Positive and negative control, the lac operon, identification of operator and regulator sequences by mutations, induction and repression, Foot-printing and gel-shift assays for identification of protein-DNA interactions. Catabolite repression. *Trp* operon – Attenuation, alternative secondary structures of *trp* mRNA.

Regulation of gene expression in eukaryotes- Response elements, DNA-binding motifs, steroid receptors, association of methylation and histone acetylation with gene expression.

Units V

Post transcriptional modifications in eukaryotes- RNA processing- mRNA 5’ capping and 3’poly-adenylation, introns and exons, RNA splicing,- spliceosome assembly, alternative splicing, processing of tRNA and rRNA, self-splicing, ribozymes, RNA editing- substitution and insertion/deletion editing, Genome editing-CRISPR- Cas technology

Post translational modification of proteins- Proteolytic cleavage, covalent modifications, glycosylation of proteins, disulfide bond formation, Protein sorting – signal peptides, transport of secretory proteins, Golgi and post-golgi sorting, coated vesicles, targeting of mitochondrial,

lysosomal and nuclear proteins, Protein degradation-Ubiquitination of proteins, Protein folding-chaperones

Reading List (Print and Online)

1. Molecular Biology Free Online Course by MIT Part 3: RNA Uploaded by edX
2. https://mooc.es/course/molecular-biology/
3. https://onlinecourses.swayam2.ac.in/cec20_ma13/preview
4. https://learn.genetics.utah.edu/
5. https://www.cellbio.com/education.html
6. https://lifescienceinteractive.com/category/molecular-biology/

Self-Study

. 1. Multiple roles of noncoding RNAs (long ncRNA ,siRNA, miRNA) in development and differentiation; implication of ncRNAs in pathologies.

2. mRNA degradation- nonsense-mediated decay.

Recommended Texts

1. Lewin's Genes XII : 12th edition, Krebs JE, Goldstein ES, Kilpatrick ST ;Prentice Hall, Delhi
2. Molecular Biology of the Gene : 6th edition, Watson JD , Baker TA, Bell S, Gann A, Levine M, Losick R; Cold Spring Harbor Laboratory Press, New York
3. Essential Cell Biology :3rd edition, Alberts B, Bray D, Hopkin K, Johnson A, Lewis J, Raff M, Roberts K, Walter P ; Garland Science, New York
4. Molecular Cell Biology : 8th edition , Lodish H, Arnold Berk; W.H.Freeman & Co, New York
5. Karp's Cell and Molecular Biology: Concepts and Experiments, 8th Edition; Wiley, India
6. An Introduction to Genetic Analysis 12th edition,, Griffith A. F, Doebley J, Peichel C, David A, Wassarman DA; Albion Press.W.H.Freeman & Co ,New York

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4)- Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	S	S	L	M	L	S	S	S	S
CO 2	S	S	S	M	M	L	M	S	S	S
CO 3	S	S	S	L	M	L	M	S	S	S
CO 4	S	S	S	M	M	L	S	S	S	S
CO 5	S	S	S	S	S	M	M	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215AEC33	Gene Editing, Cell and Gene therapy	4	1	0	4

Pre-requisites, if any: To introduce students molecular basis of cell gene therapy; viral and nonviral gene transfer techniques and gene therapy applications in hereditary and acquired diseases.

Course Objectives

1. To train the student in techniques related to the molecular basis of genetic diseases and to incorporate skills essential for various types of sequencing.
2. To inculcate practical knowledge on comparing the animal models used to model genetic diseases
3. To introduce and also elaborate knowledge about wide varieties of vectors and their features in addition to their applications and to identify the viral and nonviral gene transfer techniques
4. To educate about the characteristics of cell culture, therapeutic strategies in gene therapy with relevant safety/ethics involved and patents as well.

Course Outcomes After completion of the course, the students should be able to:

CO1. Ability to read, and evaluate scientific articles within the subjects of immune therapy, gene therapy and cell therapy. (K1, & K2)

CO2. To clone gene of their interest for several downstream purposes with a robust comprehension about wide variety of applicable gene delivery vectors. (K1, K2 & K5)

CO3. Be able to provide examples of diseases that can be treated with immune therapy, gene therapy and cell therapy. (K2, K3 & K4)

CO4. To identify knowledge gaps and need for further research within their chosen topic of immune therapy, gene therapy or cell therapy. (K2, K4 & K5)

CO5. To critically discuss and reflect on ethical and social aspects of using immune, gene or cell therapy. The student will be persuaded to contemplate on upcoming technologies for futuristic benefits. (K2, K5 & K6)

Units I Gene Editing: Basis of gene editing, DNA repair mechanisms, Double strand DNA breaks, Nonhomologous End-Joining (NHEJ), Homology directed repair, Programmable nucleases for gene editing, Meganucleases, Zinc-Finger nucleases, Transcription Activator-Like Effector Nucleases (TALEN), CRISPR-Cas systems, gene editing using CRISPR-Cas, drawbacks and major challenges to present gene editing techniques, gene editing for human disease therapy

Units II Gene and cell therapy: Basics of Gene and cell therapy, types of gene therapy, gene therapy strategies, therapeutic targets for gene therapy, choice of the therapeutic target, administration routes, delivery systems, expression of transgene, persistence of the gene therapy, cell targeting, immunological response to the therapy, ethical and legal issues, concerns about gene and cell therapy

Units III	Vectors for Gene therapy: Non-viral and viral vectors for gene therapy, Physical methods of gene delivery, Polymer, Lipid and inorganic material based chemical systems for gene delivery, Viral vectors, Lentiviral, Adenoviral, Adeno-associated virus, Herpes Simplex virus, vaccinia, baculoviral vectors for gene delivery, choice of viral vector and oncolytic virus. Gene therapy applications, Gene therapy for cancer, suicide and oncolytic gene therapy.
Units IV	Stem cells and tissue regeneration: Adult and fetal stem cells, embryonic stem cells, cell reprogramming, induced pluripotent stem cells (iPSC), Chemically induced pluripotent stem cells (CiPSC), reprogramming factors, iPSC derived progenitors 'cells, Organoids, three dimensional (3D) bioprinting.
Units V	Regulatory and Ethical Considerations of stem cell and Gene Therapy, pluripotent stem cell-based cell replacement therapies. Assessing Human Stem Cell Safety, Use of Genetically Modified Stem Cells in Experimental Gene Therapies. Technological challenges towards development of pluripotent stem cell-based cell replacement therapies.
Reading List (Print and Online)	1. Stem Cell Biology, Daniel Marshak, Richard L. Gardener and David Gottlieb, Cold Spring Harbour Laboratory Press 2. Stem cell biology and gene therapy, Booth C., Cell Biology International, Academic Press 3. Stem Cell and Gene-Based Therapy: Frontiers in Regenerative Medicine, Alexander Battler,
Self-Study	1. Applications of gene editing strategies 2. CART therapy for Cancer
Recommended Texts	1. An Introduction to Human Molecular Genetics (2nd Edition), J.J. Pasternak, 2005 2. An Introduction to Molecular Medicine and Gene Therapy 1st Edition by Thomas F. Kresina Upadhyay, S. K. (Ed.). (2021). 3. Human Molecular Genetics (4th Edition), Tom Strachan & Andrew Read, 2010.

4. Stem Cells Handbook: Stewart Sell, Humana Press; Totowa NJ, USA; Oct. 2003,

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Observe, Explain.

Analyse (K4)- Finish procedure in stepwise manner, Differentiation between various ideas, Map knowledge

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons.

Create (K6)- Check knowledge in specific or offbeat situations, Discussion, Debating, Presentation

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	L	M	S	M	M	M	M	M	M
CO 2	S	S	S	S	M	M	M	M	M	S
CO 3	S	M	S	S	M	S	S	S	S	S
CO 4	S	L	M	M	M	M	S	M	M	S
CO 5	S	S	S	S	S	S	S	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215SEC34L	Clinical Biochemistry Lab	0	0	4	4

Pre-requisites, if any: Knowledge on basic principles, Instrumentation of Biochemical techniques and metabolic reactions

Course Objectives

1. To instill skill in students enabling them to apprehend the wider knowledge about principles and techniques to be employed for the investigation of biological samples, clinical approach, normal values of biochemical constituents and clinical interpretations.
2. To inculcate the knowledge of collection, preservation of blood sample and learning various hematological parameters and their significance.
3. To perform experiments to assess liver functions. And also to study the marker enzymes of liver
4. To evaluate lipid profile and assess their relation to cardiac function.
5. To perform experiments to estimate blood glucose and glycosylated hemoglobin.
6. To perform urine analysis, estimate BUN and clearance test to assess renal function .
7. To learn basic immuno technniques antigen –antibody reactions.
8. To perform data analysis in using MS Excel
9. To introduce visit to hospital so that students may be aware of Phleobotomy ,Collection and storage of specimen, Good laboratory practices, Automation and current methods adopted in the diagnostic labs

Course Outcomes After completion of the course, the students should be able to:

CO1. The student will be able to acquire knowledge and skill in hematology techniques. They will get familiar with methods and knowledge to interpret the electrolyte concentration in serum

(K1,K2,K3,K4,K5)

CO2. The student will be able to assess the Liver Function and interpret the biochemical investigation in a given clinical situation (K1,K2,K3,K4,K5)

CO3.Skill to perform the Renal function test to assess the function of Kidney and report the abnormal parameters with reference range will be achieved by the student (K1,K2,K3,K4,K5)

CO4. To estimate the blood glucose content and lipid profile , to evaluate the alterations and record the observation in accordance to reference range will be acquired by the student (K1,K2,K3,K4,K5,K6)

CO5: The Group Experiments will support them to acquire practical skills to work in health care sector and assist them to understand the automation process in clinical labs (K1,K2,K3,K4,K5,K6)

I Haematology:

RBC count, WBC count – total and differential count, ESR, PCV, MCV. Bleeding Time, Clotting Time and Estimation of hemoglobin.

Determination of Electrolytes :Sodium, Potasium and Calcium

II Liver function test:

Estimation of bilirubin – direct and indirect. Estimation of plasma protein, A/G ratio, Thymol turbidity test, Prothrombin Time (PT),Assay of serum glutamate oxaloacetate transaminase, alkaline phosphatase, Gamma-glutamyltransferase (GGT), isoenzyme separation of LDH by electrophoresis.

III Renal function test:

Collection and Preservation of Urine sample

Qualitative tests for normal and pathological components of urine.

BUN: Estimation of blood Urea, creatinine, and uric acid.

Urea Clearance test

IV Estimation of blood glucose by orthotoluidine and glucose oxidase method. Determination of glycosylated Hb. Glucose tolerance test. Kit

	method Lipid profile: Estimation of cholesterol by Zak's method, lipoprotein profile, estimation of ketone bodies, estimation of triglycerides, free fatty acids and phospholipids.
V	Group Experiments a. Antigen – Antibody Reaction - HCG kit method , RA kit method b. Phlebotomy –Venipuncture , Different techniques of venipuncture c. Collection of blood ,Serum or Plasma separation and Storage d. Automation in Clinical Biochemistry -Autoanalyser ,Semiautoanalyser
Reading List (Print and Online)	1. https://www.researchgate.net/publication/260182512_ Practical_Manual_in_Biochemistry_and_Clinical_Biochemistry 2. https://main.icmr.nic.in/sites/default/files/upload_documents/ GCLP_Guidelines_2020_Final.pdfhttps://www.westgard.com/clia.html 3. https://www.researchgate.net/publication/263929434_Biochemistry 4. https://ucms.ac.in/Lectures-C-2020/Renal%20function%20Tests%20-%20PPT.pdf 5. https://youtu.be/i2PfjEks4GQ 6. https://www.euro.who.int/__data/assets/pdf_file/0005/268790/WHO-guidelines-on-drawing-blood-best-practices-in-phlebotomy-Eng.pdf
Self-Study	1. Laboratory handling of human biological specimen 2. Automation in Clinical Biochemistry
Recommended Texts	1 Practical Clinical Biochemistry- Varley's by Alan H Gowenlock, published by CBS Publishers and distributors, India Sixth Edition ,1988. 2. Manipal Manual of Clinical Biochemistry (For Med.Lab.And Msc Stud.) 2013 (4 Edition) 3. Case Oriented Approach in Biochemistry-Dr. Rajesh Kawaduji

Jambhulkar, Dr. Abhijit D. Ninghot: 2019 First Edition

4. Medical Lab Technology Vol I& II, Kanai L Mukerjee New Delhi: Tata Mcgraw Hill Publishing Company, 1996.
5. Practical Biochemistry – Plummer, New Delhi: Tata Mcgraw Hill Publishing Company, 2000.
6. Introductory practical Biochemistry – S.K. Sawhney, Randhir Singh, 2nd ed, 2005.

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create(K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	**S**	**S**	**S**	**S**	**M**	**S**	**L**	**S**	**M**	**S**
CO 2	**S**	**S**	**S**	**S**	**M**	**S**	**L**	**S**	**M**	**S**
CO 3	**S**	**S**	**S**	**S**	**M**	**S**	**M**	**S**	**M**	**S**
CO 4	**S**	**S**	**S**	**S**	**M**	**S**	**M**	**S**	**S**	**S**
CO 5	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**	**S**

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC35**A**	Biostatistics and Data Science	4	1	0	3

Pre-requisites, if any: Basic knowledge of Statistics and Computer Applications

Course Objectives

1. To summarize the data and to obtain its salient features from the vast mass of original data.
2. To understand the concept of various measures of dispersion.
3. To understand the concepts of sampling and learning test of significance.
4. To understand the concept of various attributes and relate to biological studies.
5. To gain knowledge in SPSS, a software package which gives a perfect graphical representation and appropriate result for the data that has been entered

Course Outcomes After completion of the course, the students should be able to:

CO1: Concepts of statistical population and sample, variables and attributes. Tabular and graphical representation of data based on variables.(K1,K2,K3)

CO2: Conditions for the consistency' and criteria for the independence of data based on attributes. Measures of central tendency, Dispersion, Skewness and Kurtosis.(K1,K2,K3)

CO3:Learning different sampling methods and analysing statistical significance.(K1,K2,K3,K4)

CO4: Understanding students t test , ANOVA , Chi square test to

analyse the significance of various research. (K1,K2,K3,K4)

CO5: Learning on data science, algorithm for machine learning, artificial intelligence and big data, their applications in clinical and pharma domain . (K1,K2,K3,K4.K6)

Units I Nature of biological and clinical experiments – Collection of data in experiment- Primary and secondary data. Methods of data collection. Classification and tabulation. Different forms of diagrams and graphs related to biological studies. Measures of Averages- Mean, Median, and mode. Use of these measures in biological studies.

Units II Measures of Dispersion for biological characters – Quartile deviation, Mean deviation, Standard deviation and coefficient of variation. Measures of skewness and kurtosis. Correlation and regression – Rank correlation – Regression equation. Simple problems based on biochemical data.

Units III Basic concepts of sampling- Simple random sample stratified sample and systemic sampling. Sampling distribution and standard error. Test of significance based on large samples. Test for mean, difference of means, proportions and equality of proportions.

Units IV Small sample tests – Students‘t’ test for mean, difference of two way means, tests for correlation and regression coefficients. Chi-square test for goodness of a non independence of attributes. F test for equality of variances. ANOVA- one way and two way. Basic concept related to biological studies

Units V Introduction to Data Science, Definition of data science, importance, and basic applications, Machine Learning Algorithms, Deep Learning, Artificial Neural Networks and their Application, Reinforcement Learning, Natural Language Processing Artificial Intelligence (AI), Data Visualization, Data Analysis, Optimization Techniques, Big Data,

Predictive Analysis. Application of AI in medical, health and pharma industries.

Reading List (Print and Online)	1.https://www.ibm.com/docs/en/SSLVMB_28.0.0/pdf/Accessibility.pdf 2.https://pure.tue.nl/ws/portalfiles/portal/19478370/20160419_CO_Mzolo.pdf 3.https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5453888/ 4.https://home.ubalt.edu/ntsbarsh/excel/excel.htm 5.https://students.shu.ac.uk/lits/it/documents/pdf/analysing_data_using_spss.pdf 6.https://www.ibm.com/support/pages/ibm-spss-statistics-28-documentation
Self-Study	1.Simple problems on probability, theoretical distributions, hypothesis testing 2. Relationship between mean, median and mode pros and cons of the measures of central tendency and deviation
Recommended Texts	1. Zar, J.H. (1984) "Bio Statistical Methods", Prentice Hall, International Edition 2. Sundar Rao P. S.S., Jesudian G. & Richard J. (1987), "An Introduction to Biostatistics", 2nd edition,. Prestographik, Vellore, India,. 3. Warren,J; Gregory,E; Grant,R (2004), "Statistical Methods in Bioinformatics",1st edition,Springer 4. Milton,J.S.(1992),. "Statistical methods in the Biological and Health Sciences", 2nd edition ,Mc Graw Hill, 5. Rosner,B (2005), "Fundamentals of Biostatistics", Duxbury Press 6. Introducing Data Science, Davy Cielen, Anro DB Meysman, Mohamed Ali.

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain

Analyse (K4) - Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create (K6) - Check knowledge in specific or off beat situations, Discussion, Presentations

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	S	S	S	M	S	L	S	S	S
CO 2	S	S	S	S	M	S	L	S	S	S
CO 3	S	S	S	S	S	S	M	S	S	S
CO 4	S	S	S	S	S	S	M	S	S	S
CO 5	S	S	S	S	S	S	M	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC35B	Immunology	5	0	0	4

Aim:

To learn the immune system and reaction

Objectives:

- To expose the students with the immune system of human body

Outcomes:

- CO1 The students may understand the immune system, its components and various techniques used in bio manipulation.
- CO2 The course will provide technical knowledge as to how different diseases are caused and various responses mediated by living cells to combat pathogen attack.
- CO3 Compare and contrast the origin, maturation process, and general function of B and T lymphocytes.
- CO4 At The course will provide sound knowledge of how immune system deals with various pathogens, different processes and cell types involved in prevention of disease.
- CO5 To understand the principles of tolerance, autoimmunity and the role of immunity in protection against pathogens.
- CO6 Along with this the students will become aware about concept, synthesis and action mechanism of vaccines.

UNIT-I

Infection

Types – Factors influencing infection – endotoxins and exotoxins – pathogenicity and virulence – sources of infection agents – carriers – portals of entry. **Immunity:** Innate – Acquired – Active and Passive Immunity – phagocytosis – Inflammation. **Immune system:** Components – Lymphoid organs, Primary, Secondary, Tertiary – Lymphoid Tissues – Cells of the immune system.

UNIT-II

Antigens: Antigenicity – Immunogenicity – Types of antigens – Haptens – specificity – Blood group antigens – Blood grouping. **Immunoglobulins:** Isolation – structure and function – Antibodies – clauses of immunoglobulins – Biosynthesis and antibody diversity. **Complement:** Complement – chemical and alternate pathway – Consequence of activation – Anaphylotoxins.

UNIT-III

Immune response: B and T cell development Cellular interaction – APC – MHC – cytokines – TCR – cluster of differentiation – HI and CMI – Regulation of immune response – Primary and secondary immune response – Immune tolerance - Immune suppression.

UNIT-IV

Immunological Techniques: Antigen – Antibody reaction – Precipitation – Agglutination – CTI – Torsion neutralization – Immunodiffusion – Immunofluoroscence – Immune adherence – Immunoelectrophoresis – EIA – ELISA – RIA – Immunochemetology – Hybridoma Technology – Merits and Demerits – Production of Vaccines and their uses. Production of antisera – Fractionation of leucocytes – Identification of lymphocytes and their subsets – Experimental animal models – Inbred strains – SCID Mice , Nude mice – mice cell culture system.

UNIT-V

REFERENCE:

1. Immunology – Richard.A.Goldshy and Kuby.
2. Essential Immunology – Roitt et al.
3. Immuno biology – Janeway. C: Paul Travers.
4. Immunology, Short Course – Eli Benjamin AV. et al.
5. Fundamentals of Immunology Springer Verlag – Wier et al.
6. A Handbook of Practical Immunlogy – G.P.Talwar.
7. Fundamental Immunology – Coleman.
8. Textbook of microbiology – Anantha marayanan & panikar.

Course Code	Course Title	L	T	P	C
23215SEC36	Molecular basis of disease and therapeutic strategies	4	1	0	3

Pre-requisites, if any: Knowledge of Human Physiology, Metabolism and Clinical Biochemistry

Course Objectives

1.To understand the concepts of the mechanisms involved in regulation of blood sugar and management of diabetes mellitus

2.To gain in-depth knowledge of the mechanisms of cancer and of tumor metastasis

3.The student will review the basic organization of the central and peripheral nervous system that coordinate the sensory and motor functions of the body. In addition, the student will explore impaired features underlying the major neuropathological complications.

4.To gain knowledge in renal diseases

5.To understand the mechanisms involved in cardiac disorders

Course Outcomes

On completion of this course the student will be able to understand

CO1. Overall view about the complications of diabetes mellitus and its management.

CO2. Comprehensive understanding of the concepts of cancer biology and implicating the theoretical concepts for further research

CO3.Understand and appreciate the pathophysiology of conditions affecting the nervous system.

CO4.A thorough knowledge of renal and cardiac diseases with emphasis related to mechanistic aspects and therapeutic interventions.

CO5. A thorough knowledge on the experimental models of non-communicable diseases that will be applied for future research or project dissertation. An in-depth knowledge on development of drugs against non-communicable diseases.

Units

I Mechanism of blood sugar regulation in human body. Pathophysiology of Type I and II diabetes, Diabetes – investigation methods for the diagnosis of diabetes. Nutritional care. Complications related to diabetes – Diabetic cardiovascular disease, retinopathy, neuropathy and nephropathy. Cellular and molecular mechanism of development of diabetes- Management of Type I and Type II diabetes, drugs for the treatment of diabetes.

II Biology of cancer: Overview of hallmarks of cancer. Tumorigenesis, Tumor progression and mechanism of Metastasis. Proto-oncogene to oncogene. Oncogene- myc and src family. Tumor suppressor gene-Rb and p53 pathway in cancer. Diagnosis- Non-invasive imaging techniques, Tumor diagnosis, Interventional radiology, New imaging technique, Molecular techniques in cancer diagnosis.- treatment of cancer- surgery, radiotherapy, chemotherapy, hormonal treatment, and biological therapy. Introduction to personalized medicine.

III Brain- neuronal network- memory- Neurogenerative diseases- Parkinson and Alzheimer Disease- molecular understanding of the neurodegenerative diseases- treatment modalities.

IV Acute and chronic renal failure, glomerular diseases–

	glomerulonephritis, nephritic syndrome, diabetes insipidus, diagnosis of kidney disease.
V	Introduction to cardiovascular diseases, Lipids and lipoproteins in coronary heart disease-cardiac enzymes, Molecular changes during cardiac remodeling – hypertrophy of hearts – heart failure- treatment modalities.

Reading List (Print and Online)

1. The Biochemical basis of disease:2018, **Barr AJ;** Portland Press
2. Biochemical Basis of Diseases
3. https://www.biologydiscussion.com/diseases-2/biochemical-basis-of-diseases/44276

Recommended Texts

1. Wills' Biochemical Basis of Medicine: 2nd edition, Thomas H, Gillham B;Elsevier
2. Molecular Biochemistry of Human Diseases,2021, Feuer G ,de la Iglesia F; CRC Press

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview

Application (K3) - Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain

Analyse (K4) - Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	**S**	**S**	**S**	**M**	**M**	**S**	**S**	**S**	**S**	**S**
CO 2	**S**	**M**	**S**	**L**	**M**	**M**	**M**	**M**	**M**	**S**
CO 3	**S**	**S**	**M**	**L**	**S**	**S**	**M**	**M**	**S**	**M**
CO 4	**S**	**M**	**M**	**M**	**M**	**M**	**S**	**S**	**M**	**S**
CO 5	**S**	**S**	**M**	**M**	**S**	**M**	**M**	**M**	**S**	**S**

S-Strong M-Medium L-Low

SEMESTER IV

Course Code	Course Title	L	T	P	C
23215AEC41	Pharmaceutical Biochemistry	4	1	0	4

Pre-requisites, if any: The student should have a basic knowledge of drug discovery and development. Student should possess basic knowledge bioinformatics to understand and correlate the drug development process.

Course Objectives

1. To understand the different types of bioinformatic tools for drug discovery.
2. To get an overview of how different bioinformatic tools aid in the process of target identification, drug screening and quantitative structure activity relationship.
3. To assimilate the involvement of different metabolic pathways involved in drug metabolism and correlate their involvement in elimination process
4. To understand the biochemical basis of drug action at the target tissue.
5. To understand different phases in drug clinical trials and its assessment.

Course Outcomes

After completion of the course, the students should be able to:

CO1. To understand and explain the basic concepts of drug discovery and drug development process.

CO2. To review the different software and computational tools which aid in the design of drugs and its rationalization.

CO3. To analyze the different stages of the drug discovery process with the target & hit identification, assays for drug screening and preclinical studies.

CO4. To understand the various phases of the clinical trails and the method of conduct of clinical trails.

Units

I Drug discovery and development, drug target identification and validation, Hit identification, General principles of screening, correlations between various animal models and human situations, Correlation between in-vitro and in-vivo screens; Special emphasis on cell-based assay, biochemical assay, radiological binding assay, Pharmacological assay, In vitro, In vivo & Ex-vivo experiments, lead optimization, preclinical studies.

II Bioinformatics approaches for drug development:

Identification of potential molecules, chemical compound library preparation, Identification of target in pathogen, Ligand & protein preparation, Molecular docking, Binding free energy estimation, High throughput virtual screening, Docking protocol validation and enrichment analysis, Single point energy calculation, Pharmacokinetics and Pharmacodynamics, ADME & toxicity prediction, Molecular dynamic simulation, Rule of three and five, Lipinsky rule, Pharmacophore development, Quantitative structure activity relationship, 3D-QSAR, Techniques of developing a pharmacophore map covering both ligand based and receptor based approaches.

III Drug metabolism & interactions:

Drug-receptor interactions, receptor theories and drug action, Xenobiotics, xenobiotics phases (Phase-I, Phase-II and Phase-III), role of cytochrome P450 oxidases and glutathione S-transferases in drug metabolism, factors affecting drug metabolism, Enzymes as a drug target, Kinase inhibitors, ATPase inhibitors, drug protein interaction, DrugDNA interaction. Basic ligand concepts-agonist, antagonist, partial agonist, inverse agonist, efficiency and potency. Forces involved in drug-receptor complexes. Receptor classification – the four super families. Receptor binding

assays- measurement of Kd, Bmax and IC_{50}.

IV	Biochemical mode of action of antibiotics- penicillin and chloramphenicol, actions of alkaloids, antiviral and antimalarial substances. Biochemical mechanism of drug resistance- sulphonamides. Drug potency and drug efficacy. General principles of chemotherapy: chemotherapy of parasitic infections, fungal infections, viral diseases. Introduction to immunomodulators and chemotherapy of cancer.
V	Clinical trials (Phase-I, Phase-II, Phase-III and Phase-IV clinical trial). Main features of clinical trials, including methodological and organizational considerations and the principles of trial conduct and reporting. Key designs surrounding design, sample size, delivery and assessment of clinical trials.
Self-Study	1. Examples of pharmaceutical development of a drug 2. Basic pharmacology of drug action and kinetics
Reading List (Print and Online)	1. Textbook of Drug Design. Krogsgaard-Larsen, Liljefors and Madsen (Editors), Taylor and Francis, London UK, 2002. 2. Drug Discovery Handbook S.C. Gad (Editor) Wiley-Interscience Hoboken USA, 2005
Recommended Texts	1. Practical Application of Computer-Aided Drug Design, Ed. Charifson P., Marcel Dekker Inc. 2. 3D QSAR in Drug Design: Theory, Methods and Applications, Ed. Kubinyi H., Ledien 3. Pharmaceutical Profiling in Drug Discovery for Lead Selection, Borchardt RT, Kerns, EH, Lipinski CA, Thakker DR and Wang B, AAPS Press, 2004 4. Drug Discovery and Development; Technology in Transition. HP Rang. Elsevier Ltd 1st edition 2006.

5. Pharmacology in Drug Discovery. T. P. Kenakin. Elsevier, 1st Edition 2012.

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create(K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	M	S	M	S	M	M	S	S	S
CO 2	S	S	S	M	M	S	S	S	S	S
CO 3	S	S	S	L	S	M	M	S	S	M
CO 4	S	M	S	L	S	L	M	S	S	M
CO 5	S	S	S	L	S	M	M	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215AEC42	Biochemical Toxicology	4	1	0	4

Pre-requisites, if any: The student should have a basic knowledge of pharmacology of drug action and understanding on their biochemical pathways.

Course Objectives

1. To understand the detailed study of biochemical basis of drugs and its toxicity, particularly their actions on living systems.
2. To understand the relevance and methods to identify the chemotherapeutic value of drug.
3. To understand the fundamentals of toxicology and dose- response relationships.
4. To understand the toxicological drug testing procedures based on in vitro and animal studies
5. To understand biochemical pathways of drug toxicity and its manifestation on vital organs.

Course Outcomes On completion of this course, the student will be able

CO1: To appreciate and understand the role of toxicological biomarkers to assess drug toxicities.

CO2: To conceive the role of disposition of drug in human system and their metabolism and methodologies pertaining to toxicological studies.

CO3: To understand and evaluate the functions of different organs on drug disposition and associated drug toxicities.

CO4 : To understand the toxicological response to foreign compounds and their pharmacological, physiological and biochemical effects.

CO5: To link the mechanism of toxicity and clinical symptoms with underlying physiological disturbances.

Units

I	Fundamentals of Toxicology and dose-Response Relationships: Introduction Biomarkers Criteria of Toxicity New Technologies Evaluation of Toxicity Interactions; Dose Response; Measurement of Dose-Response; Relationships Linear Dose Response Hormesis; Hazard and Risk Assessment Duration and Frequency of Exposure and Effect
II	Factors Affecting Toxic Responses: Disposition: Absorption ,Sites of absorption, distribution, Excretion; Metabolism: types of Metabolic change phase I reactions; Phase 2 reactions; control of Metabolism, Toxication vs. Detoxication
III	Toxicity testing; Test protocol, Genetic toxicity testing & Mutagenesis assay: In vitro test systems: bacterial mutation tests-Reversion test,Ames test, Fluctuation test, and Eukaryotic mutation test. In vivo test system Mammalian mutation test-Host mediated assay and Dominant Lethal test. Biochemical basis of toxicity: Mechanism of toxicity: Disturbance of excitable membrane function, Altered Calcium homeostasis, Covalent binding to cellular macromolecules & genotoxicity, Tissue specific toxicity
IV	Toxic Responses to Foreign Compounds: Direct Toxic Action: Tissue Lesions; Mechanism and response in cellular toxicity, pharmacological, physiological and Biochemical effects; Developmental Toxicology-Teratogenesis; Immunotoxicity Genetic Toxicity; Chemical Carcinogenesis
V	Biochemical Mechanisms of Toxicity: Tissue Lesions: Liver Necrosis; kidney Damage; Lung Damage, Liver damage, Cardiac damage;

	Neurotoxicity; Exaggerated and Unwanted pharmacological effects; Physiological effects; Biochemical Effects: Lethal Synthesis and Incorporation, Interaction with specific Protein Receptors; Teratogenesis; Immunotoxicity; multi-Organ Toxicity
Self-Study	• Case studies to review
Reading List (Print and Online)	1. Preclinical Safety Evaluation of Biopharmaceuticals: A Science-Based Approach to Facilitating Clinical Trials by Joy A. Cavagnaro 2. A Comprehensive Guide to Toxicology in Nonclinical Drug Development 2nd Edition by Ali S. Faqi
Recommended Texts	1. Principles Of Toxicology by: Karen E Stine, Thomas M Brown 2006 Publisher. Crc Press 2. Principles of Biochemical Toxicology by John A. Timbrell Publisher: Informa Healthcare 3. Environmental Toxicology by Sigmund F. Zakrzewski, (2002) Publisher: Oxford University Press, USA

Method of Evaluation:

Test I	Test II	Assignment	End Semester Examination	Total	Grade
10	10	5	75	100	

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create(K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	S	S	L	S	L	M	M	M	M
CO 2	M	M	S	M	M	L	M	S	S	S
CO 3	S	S	S	M	M	L	S	S	M	M
CO 4	S	M	S	M	M	M	S	S	M	M
CO 5	M	S	S	S	S	M	M	M	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC44A	Biosafety, Lab Safety and IPR	4	1	0	3

Pre-requisites, if any: The student should have a basic knowledge of hazards associated with the handling of biological agents and importance of intellectual property from scientific research.

Course Objectives

1. To assimilate the hazards associated with the handling of biological and chemical agents.
2. To understand how to protect from the hazards by the implementation of various safety measures in biochemical laboratories.
3. To implicate the importance of protecting the scientific intellect by filing patent and understand the various offices for filing and maintaining patents
4. To understand the scope of patenting in biological research.
5. To create an awareness of ethics associated with used of genetically modified organisms/cells and its rationale for use in living organisms.

Course Outcomes After completion of the course, the students should be able to:

CO1. To understand and implement various aspects of biosafety and carry out risk assessment of products in biological research

CO2. Understand the basic concepts of ethics and safety that are essential for different disciplines of science and procedures involved and protection of intellectual property and related rights.

CO3. To appreciate the intellectual property rights and its implementation of on the invention related to biological research.

CO4. To understand the statutory bodies that regulate the property rights and its validity in various countries.

CO5. Critique the ethical concerns associated with modern

biotechnology processes and plan accordingly.

Units

I Biosafety: Historical background; introduction to biological safety cabinets; primary containment for biohazards; biosafety levels; recommended biosafety levels for infectious agents and infected animals; biosafety guidelines - government of India, roles of IBSC, RCGM, GEAC etc. for GMO applications in food and agriculture; environmental release of GMOs; risk assessment; risk management and communication; national regulations and international agreements.

II Laboratory safety - Chemical, electrical and fire hazards; handling and manipulating human or animal cells and tissues, toxic, corrosive or mutagenic solvents and reagents; mouth pipetting, and inhalation exposures to infectious aerosols, Safe handling of syringe needles or other contaminated sharps, spills and splashes onto skin and mucous membranes. Health aspects; toxicology, allergenicity, antibiotic resistance.

History of biosafety microbiology and molecular biology, Risk assessment, Personal protective equipment, Laboratory facilities and safety equipment, Disinfection, decontamination, and sterilization, Regulatory compliance, Laboratory security and emergency response and administrative controls.

III Intellectual Property Rights (IPR): Introduction to patents, types of patents, process involved in patenting in India, trademarks, copyright, industrial design, trade secrets, traditional knowledge, geographical indications, history of national and international treaties and conventions on patents, WTO, GATT, WIPO, Budapest Treaty, Patent Cooperation Treaty (PCT) and TRIPS. Patent databases: Searching international databases; analysis and report formation. Indian Patent Act 1970; recent

	amendments; filing of a patent application; precautions before patenting disclosure/non-disclosure; procedure for filing a PCT application. The patentability of microorganisms-claims, Characterization and repeatability disposition in the culture collections, legal protection for plants and other higher organisms, new plant varieties by rights, tissue culture protocols
IV	Patent filing and infringement: Patent application- forms and guidelines, fee structure, time frames; types of patent applications: provisional and complete specifications; PCT and convention patent applications, International patenting-requirement, financial assistance for patenting-introduction to existing schemes; Publication of patents-gazette of India, status in Europe and US. Research Patenting: Patenting by researchers and scientists-University/organizational rules in India and abroad. Detailed information on patenting biological products, Case studies on patents (basmati rice, turmeric, neem etc.), and patent infringement.
V	Bioethics: Introduction to bioethics, human genome project and its ethical issues, genetic manipulations and their ethical issues, ethical issues in GMOs, foods and crops in developed and developing countries, environmental release of GMOs, ethical issues involved in stem cell research and use, use of animals in research experiments, animal cloning, human cloning and their ethical aspects, testing of drugs on human volunteers.
Self-Study	1. Review of drug patent documents 2. Safety in biological research laboratories
Reading List (Print and Online)	1. V. Shree Krishna, (2007). Bioethics and Biosafety in Biotechnology, New Age International Pvt. Ltd. Publishers. (Unit III, Unit IV and Unit V) 2. Deepa Goel, Shomini Parashar, (2013). IPR, Biosafety and Bioethics, Pearson. (Unit II) 3. R. Ian Freshney, 2016. Culture of Animal Cells: A Manual of Basic

Technique and Specialized Applications, 6th Ed, John Wiley & Blackwell.

4. BAREACT, Indian Patent Act 1970 Acts & Rules, Universal Law Publishing Co. Pvt. Ltd., 2007. (Unit I)

Recommended Texts

1. Biosafety in Microbiological and Biomedical Laboratories, (2020) 6th Ed. (https://www.cdc.gov/labs/pdf/SF__19_308133-A_BMBL6_00-BOOK-WEB-final3.pdf)
2. Kankanala C., (2007), Genetic Patent Law & Strategy, 1st Edition, Manupatra Information Solution Pvt. Ltd.,

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create(K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	S	S	M	S	M	S	S	S	S
CO 2	S	S	S	L	M	M	S	S	S	S
CO 3	S	M	M	M	S	M	S	S	S	M
CO 4	S	M	M	L	S	L	S	S	S	M
CO 5	S	S	S	L	S	M	S	S	S	S

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC44B	Plant Biochemistry	4	1	0	3

Course Prerequisites or Co-requisites A background in plant biology, general biochemistry or chemistry is expected. Note that concurrent registration in any of these courses will NOT meet this requirement. Students must have completed both semesters of O-chem and a biochemistry course that covers general metabolism prior to taking this course.

Course Objectives :

This course covers biochemical processes specific to plants and is aimed to allow students to gain an understanding and appreciation of how biochemical components are synthesized and utilized by plants during growth and development and in their interactions with their environment, as well as how these processes can be manipulated. This course includes topics in photosynthesis, carbohydrates, nitrogen and lipid metabolism, specialized metabolism and plant metabolic engineering. Flux and genomics-based techniques, such as proteomics, transcriptomics and metabolomics are discussed in relation to metabolism.

Unit-I Plant cell: Structure, function and mechanisms of action of phytochromes, cryptochromes and phototropins, stomatal movement, transpiration, photoperiodism and biological clocks, plant movement. Photosynthesis: Photosynthetic apparatus, pigments of photosynthesis, Calvin cycle (C3 plants), Hatch slack (C4 plants) & CAM pathways of carbon reduction and its regulation, Structure, function and regulation of RUBISCO, Crassulacean acid metabolism in plants. Photorespiration: photorespiration pathway and significance, cyanide resistance, relationship between photosynthesis, photorespiration.

Unit-II

Phytohormones: Biosynthesis, transport, physiological effects, mode of action and signal transduction of auxins, gibberlic acid, abscisic acid, ethylene and cytokinins in germination, embryogenesis, growth and development of plant.

Unit III

Nitrogen metabolism: Nitrogen fixation, nitrogenise complex, biochemistry and genetics of nitrogen fixation and ammonium assimilation, structure of 'NIF' genes and its regulation, structural features of nitrate reductase and nitrite reductase, regulation of nitrate and sulphate assimilation.

Unit III: Secondary plant metabolites: Nature, distribution, biosynthesis and function of plant metabolites, biosynthesis of nicotine. Biochemistry of plant toxins, phytohemagglutinins, lathyrogens, nitriles, protease inhibitors, protein toxins, role of secondary metabolites in chemical defence.

Unit-IV Revised M.Sc. Biochemistry from 2019 12 Plant stress physiology: Plant stress, plant responses to abiotic and biotic stresses, salinity, water, heat, chilling, anaerobiosis, heavy metals, radiations and their impact on plant growth and metabolism, mechanisms of resistance to biotic stress and abiotic stress, antioxidative defence mechanism. Plant defence:

Unit V: Genetic basis of plant-pathogen interactions, antio R-Avr gene interactions and isolation of R genes, hypersensitive response (HR), systemic acquired resistance (SAR) and induced systemic resistance (ISR).

Books recommended

- Introduction of Plant Biochemistry, by Goodwin T. W. and E.I. Mercer, Pergamon Press, Oxford, 1983.
- Plant Physiology, 5th Edition, by Lincoln Taiz and Eduardo Zeiger, Amazon press, 2012
- Introduction of Plant Biochemistry, by Goodwin T. W. and E.I. Mercer, Pergamon Press, Oxford.
- Buchanan BB, Gruissem W & Jones RL. 2000. Biochemistry and Molecular Biology of Plants. 2nd Ed. John Wiley.
- Dey PM &Harborne JB. 1997. Plant Biochemistry. Academic Press. • Heldt HS. 1997. Plant Biochemistry and Molecular Biology. Oxford Univ.Press.

Course Code	Course Title	L	T	P	C
23215DSC45A	Developmental Biology	4	1	0	3

Pre-requisites, if any: Comprehensive Knowledge of Cell Biology

Course Objectives The candidates undertaking this course will understand the concepts of developmental biology.

1. To understand the background of developmental biology
2. To gain in-depth knowledge of various model organisms
3. To gain insight into aspects of stem cell technology
4. To gain insights into morphogenesis and oranogenesis
5. To acquire in-depth understanding of cell death mechanisms and cell fate decision

Course Outcomes

CO1.Grasp knowledge about the background of developmental biology

CO2.Gain abundant knowledge about model oraganisms and gametogenesis

CO3.Gain knowledge about stem cells and their applications in regenerative therapy

CO4.Good knowledge about organogenesis

CO5.Learn the basics of cell death mechanisms and cell fate decision .

Units

I Overview of Developmental biology: Background of Developmental biology - Principles of developmental biology –Potency, commitment, specification, induction, competence, determination and differentiation; morphogenetic gradients; cell fate and cell lineages; stem cells; genomic equivalence and the cytoplasmic determinants; imprinting; mutants and

transgenics in analysis of development.

II Model organisms

Gametogenesis – production of gametes, Formation of zygote, fertilization and early development: molecules in sperm-egg recognition in animals; embryo sac development and double fertilization in plants;cleavage, blastula formation, embryonic fields, gastrulation and formation of germ layers in animals; embryogenesis, establishment of symmetry in plants; seed formation and germination. Drosophila Developmental biology- Axis formation, Genes & mutation. *C.elegans* – Vulva formation, Axis formation.

III Regeneration Developmental Biology

Stem cells – Definition, Classification, Embryonic and adult stem cells, properties, identification, Culture of stem cells, Differentiation and dedifferentiation, Stem cellmarkers, techniques and their applications in modern clinical sciences. Three- dimensional culture and transplantation of engineered cells. Tissue engineering - skin, bone and neuronal tissues.

IV Morphogenesis & Organogenesis:Cell aggregation and differentiation in Dictyostelium; axes and pattern formation in Drosophila, amphibia and chick; organogenesis – vulva formation in Caenorhabditis elegans, eye lens formation, limb development and regeneration in vertebrates; differentiation of neurons, post embryonic development- larval formation, metamorphosis; environmental regulation of normal development; sex determination.

V Cellular senescence and Cell fate decision

Cellular senescence – concepts & Frizzled receptor in Development and disease. Diabetes and developmental biology, Cell death pathways in developments. Markers of important diseases.

Reading List (Print and Online)

Developmental Biology – Gilbert Scott

http://bgc.org.in/pdf/study-material/developmental-biology-7th-ed-sf-gilbert.pdf

Recommended Texts Developmental biology: VIII edition, **Gilbert, S**F; Sinauer Associates, Inc

Methods of assessment:

Recall (K1) - Simple definitions, MCQ, Recall steps, Concept definitions.

Understand/ Comprehend (K2) - MCQ, True/False, Short essays, Concept explanations, Short summary or overview.

Application (K3) - Suggest idea/concept with examples, Solve problems, Observe, Explain.

Analyse (K4) – Problem-saving questions, Finish a procedure in many steps, Differentiate between various ideas.

Evaluate (K5) - Longer essay/ Evaluation essay, Critique or justify with pros and cons

Create(K6) – Check knowledge in specific or offbeat situations. Discussion.

Mapping with Programme Outcomes:

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10
CO 1	S	M	M	S	S	M	L	S	S	M
CO 2	M	M	M	M	M	S	M	S	M	M
CO 3	M	M	L	M	M	S	L	S	L	L
CO 4	S	M	L	S	S	M	S	S	M	M
CO 5	S	S	M	S	L	M	M	S	M	M

S-Strong M-Medium L-Low

Course Code	Course Title	L	T	P	C
23215DSC45B	Cancer Biology	4	1	0	3

OBJECTIVES:

To enable the students to understand • Basic biology of cancer • Impact of antibodies against cancer in the human body leading to more effective treatments • Enhanced immunology based detection methods and imaging techniques • Development of cell based and cytokine based immunotherapy against cancer.

UNIT I FUNDAMENTALS OF CANCER BIOLOGY

Regulation of cell cycle, mutations that cause changes in signal molecules, effects on receptor, signal switches, tumour suppressor genes, modulation of cell cycle in cancer, different forms of cancers, diet and cancer. Cancer screening and early detection, Detection using biochemical assays, tumor markers, molecular tools for early diagnosis of cancer.

UNIT II PRINCIPLES OF CARCINOGENESIS

Theory of carcinogenesis, Chemical carcinogenesis, metabolism of carcinogenesis, principles of physical carcinogenesis, x-ray radiation-mechanisms of radiation carcinogenesis.

UNIT III PRINCIPLES OF MOLECULAR CELL BIOLOGY OF CANCER

Signal targets and cancer, activation of kinases; Oncogenes, identification of oncogenes, retroviruses and oncogenes, detection of oncogenes. Oncogenes/proto oncogene activity. Growth factors related to transformation. Telomerases.

UNIT IV PRINCIPLES OF CANCER METASTASIS

Clinical significances of invasion, heterogeneity of metastatic phenotype, metastatic cascade, basement membrane disruption, three step theory of invasion, proteinases and tumour cell invasion.

UNIT V NEW MOLECULES FOR CANCER THERAPY

Different forms of therapy, chemotherapy, radiation therapy, detection of cancers, prediction of aggressiveness of cancer, advances in cancer detection. Use of signal targets towards therapy of cancer; Gene therapy.

TOTAL : 45 PERIODS

OUTCOMES: The course would facilitate the students

• To appreciate the role of immune system in cancer • To describe self – tolerance machinery and immune surveillance • To understand the cancer microenvironment and its influence on immune cells • To have awareness on medical applications of cytokines and immune cells against cancer

TEXT BOOKS:

1. Weinberg, R.A. "The Biology of Cancer" Garland Science, 2007
2. McDonald, F etal., " Molecular Biology of Cancer" IInd Edition. Taylor & Francis, 2004.

REFERENCES:

1. King, Roger J.B. "Cancer Biology" Addison Wesley Longman, 1996. 2. Ruddon, Raymond W. " Cancer Biology" IIIrd Edition . Oxford University Press, 1995.